First Report of Human Immunodeficiency Virus Type 1 Circulating Recombinant Form 02_AG Recombinant Strain of African Origin from North India

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Abstract: Problem statement: Human Immunodeficiency Virus Type-1 (HIV-1) displays extensive genetic diversity globally which poses a serious challenge in making efficacious vaccine and developing effective therapeutic interventions. HIV-1 epidemic in India is largely driven by genetic subtype C but lately a number of reports suggest the presence of circulating recombinant forms and unique recombinant forms from different states of India. Approach: We analyzed 13 envelope sequences spanning the region from V1-V5 from three regions of North India and subjected to extensive genetic analysis. Results: We identified equal numbers (n = 6) of subtype C and CRF02_AG recombinant strain and a single subtype B-specific envelope. The extensive phylogenetic analysis of our CRF02_AG strains indicate that they are closely related to Central and Central-West African strains. The analysis of intra-subtype nucleotide distance revealed that the subtype C sequences are significantly more divergent and diverse than CRF02_AG strains (p<0.004). Conclusion: Appearance of CRF02_AG strain of African origin among commercial sex workers from Delhi and Punjab region of India is reported for the first time which needs to be monitored for its pathogenesis and its ability to recombine with the circulating strains.

Key words: Viral genotyping, molecular epidemiology, HIV-1 genes, HIV-1, CRFs in India

INTRODUCTION

First incident of CRF02_AG strain was reported from Gabon, a South Western country of Africa in the year 1988 (Delaporte et al., 1996). Subsequently it was reported from Djibouti, Nigeria, Mali, Niger and other neighboring countries (Carr et al., 1998; McCutchan et al., 1999; Cornelissen et al., 2000). It was first reported in Europe from France (Vachot et al., 2004) in the year 1992, followed by Netherlands, Sweden, Spain (mainly from the immigrants from west and central west Africa) (Holguin et al., 2000). Among the Asian countries the first report was from Republic of Korea (Kim et al., 1999). The first case of HIV/AIDS in India appeared in the year 1985. The progress of HIV epidemic has relatively been slow initially. Over the years the number of HIV infected persons have increased and reported from almost all the states of India and is a major public health problem now. The estimated number of HIV-1 infected individuals was 2.5 million in the year 2008 which placed India in the third position behind South Africa and Nigeria (UNAIDS, 2008). Global HIV-1 spread coupled with extraordinary ability of the virus to mutate poses a great challenge to the clinicians and researchers in terms of devising successful therapy and in the development of effective vaccine. In India, the predominant HIV-1 genetic subtype is C followed by Subtypes B, A and E along with several Circulating Recombinant Forms (CRFs) and Unique Recombinant Forms (URFs) co-circulating in different parts of India (Neogi et al., 2009). India is a large country and it is quite possible that isolated pockets of HIV-1 pandemic may develop independently. The geographic location of North India is important for shaping the HIV-1 epidemic because of the presence of other genetic subtypes in countries like China, Pakistan and Afghanistan. The inhabitants of these regions are
also frequent traveler to the European countries (Neogi et al., 2008).

We have analyzed partial envelope sequences from North India from four different but adjacent states (Delhi, Punjab, Haryana and Himachal Pradesh, Delhi being the capital of India). We found existence of subtypes B, C and CRF02_AG strains in this region. In this study we analyzed in detail the ancestral origin of the CRF02_AG strains from North India using phylogenetic approach and predict the possible route of spread of these strains from Africa to Europe and Asia.

**MATERIALS AND METHODS**

**Patient population and genetic characterizations:** We were able to amplify 13 Envelope sequences (V1-V5 region) from 45 enrolled patients from Delhi, Haryana, Punjab and Himachal Pradesh following all the requisite ethical clearances. The genomic DNA (pro-virus) was obtained from whole blood genomic DNA using QIAamp Blood DNA mini kit (Qiagen, Germany) and was used to amplify the envelope gene as described by us previously using nested PCR technology (Neogi et al., 2008; Bano et al., 2009). All the Polymerase Chain Reactions (PCRs) were performed with high fidelity Taq DNA polymerase (ExTaq, Takara, Japan) using the specific primers. Approximately 1.25 Kb DNA fragment corresponding to V1-V5 region was amplified initially using the following primers (FPEnv-01 and RPEnv-02). Thereafter, 700 bp fragment (V3-V5) was amplified using two internal sets of primers (FPEnv-03 and RPEnv-04). The two pairs of primer sequences were as follows:

- FPEnv-01: 5’-ATGGGATCAAAGCCTAAAGCCATGTG
- RPEnv-02: 5’-AGTGCTTCCTGCTGCTCCCAAGAACCCAAG
- FPEnv-03: 5’-CTGTTAAATGGCAGTCTAGC
- RPEnv-04: 5’-CACCTCTCCAAATTTGTCCTCCA

The cycling conditions for amplifying both the fragments were: 35 cycles at 98°C for 15 sec, 55°C for 30 sec and 72°C for 1 min with a final extension at 72°C for 10 min. PCR-amplified DNA was cloned into pGem-T (Ez) vector (Promega Biotech. WI, USA) and sequenced in both directions using T7 and SP6-specific primers. At least 4 independent clones were analyzed for each sample and only one representative clone from each sample was subjected to extensive genotypic analysis. Mother and child samples were processed separately to avoid cross contamination. It is important to mention that we were unable to amplify envelope sequences from several samples which may be due to extreme genetic variability, poor yields of genomic DNA, or due to low proviral DNA content in the cells of the infected individuals as most of them were under first line of antiretroviral treatment. The characteristics and some of the clinical parameters of 13 patients are given in Table 1.

Sequences were compared with reference strains downloaded from Los Almos database (http://www.hiv.lanl.gov). Multiple sequence analysis was performed in ClustalW 1.8.3 located in DNA data Bank of Japan (DDBJ) website (http://clustalw.ddbj.nig.ac.jp/top-e.html). The phylogenetic analysis was carried out using MEGA 4 software (Tamura et al., 2007). Genotyping was carried out using viral genotyping tools located at NCBI (http://www.ncbi.nlm.nih.gov/projects/genotyping/formage.cgi), REGA subtyping tool ver 2.0 (http://www.bioafrica.net/subtypetool/html) and RIP 3.0 (http://www.hiv.lanl.gov/content/sequence/RIP/RIP.html).

**In silico** co-receptor usage predictions were performed using Geno2pheno (http://coreceptor.bioinf.mpi-inf.mpg.de/) and WebPSSM (http://indra.mullins.microbiol.washington.edu/webpssm/) programs.

| Table 1: Characteristics of HIV-1 infected individuals (n = 13) from North India |
|-----------------------------------------------|-----------------|
| Characteristic                        | Value           |
| Age (mean ± SD) (years)              | 28.6±11.8       |
| **Gender**                          |                 |
| Male                               | 5.0             |
| Female                             | 8.0             |
| **Residence**                       |                 |
| Punjab                             | 5.0             |
| Haryana                           | 2.0             |
| Himachal Pradesh                   | 1.0             |
| Delhi                              | 5.0             |
| **Route of transmission**           |                 |
| Heterosexual                       | 10.0            |
| Vertical                           | 2.0             |
| Unknown                            | 1.0             |
| **CD4 count**                       | 480.0 (96-834)  |
| **ART status**                     |                 |
| Naive                              | 4.0             |
| Experienced                        | 9.0             |
| **Year of sampling**               |                 |
| 2007                               | 2.0             |
| 2008                               | 10.0            |
| 2009                               | 1.0             |

*: Median CD4 count is calculated from 10 patients only.
**Evolutionary analysis:** Reference sequences were downloaded from HIV-1 Los Almos Database using geographic distribution of search result tools (http://www.hiv.lanl.gov/components/sequence/HIV/search/search.html) (accessed on July 2009) and/or NCBI database from three different continents; Africa, Europe and Asia. The evolutionary history was inferred using the neighbor joining method and minimum evolution method (Rzhetsky and Nei, 1992). The evolutionary distances were computed using the maximum composite likelihood method (Tamura et al., 2004) and are in the units of the number of base substitutions per site. The ME tree was searched using the Close-Neighbor-Interchange (CNI) algorithm at a search level of 1 (Nei and Kumar, 2000). We also used UPGMA method for phylogenetic analysis. Blast search for identity was performed using NCBI Blast and HIV-1 Blast in Los Almos database.

**RESULTS**

We identified 6 subtype C, 6 CRF02_AG recombinant strains and 1 subtype B-specific envelope sequences using four different tools, namely NCBI viral genotyping tools, REGA subtyping tools, RIP 3.0 and phylogenetic analysis. Neighbor joining phylogenetic tree (Nei and Kumar, 2000) of all 13 strains are given in Fig. 1.

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**Fig. 1: Neighbor joining Phylogenetic tree of Env gene**

Note: Reference sequences of subtypes A1, B, C, D, F1, G, H, J, K, 01_AE, 02_AG and outlier group O were downloaded from Los Almos Database. The circular phylogenetic tree was constructed in MEGA 4. The Indian strains are indicated as follows: ■ subtype C; ♦ subtype B and ● CRF02_AG and their accession numbers are given. The tree was rooted with outlier group O.
Among the HIV-1 infected individuals there were 2 mother-to-child transmission cases. In both the cases the mother and child were infected with subtype C viruses and successful transmission (judged by high bootstrap value among individual mother-child pairs) was detected (data not shown). 5 CRF02_AG and 4 subtype C showed CCR5 usage. One each subtype C (Acc No. FJ769835) and subtype B (Acc. No. FJ769836) showed CXCR4 co-receptor usage (data not shown, manuscript under preparation).

Fig. 2: Evolutionary relationships of CRF02_AG strains from different continent

Note: The evolutionary history was inferred using the Neighbor-Joining method. Confidence probability was multiplied by 100 so that the interior branch length is greater than 0, as estimated using the bootstrap test (1000 replicates is shown next to the branches). The tree is drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree. The accession numbers are given followed by country initial (DJ: Djibouti, NG: Nigeria, NJ: Niger, GA: Ghana, CM: Cameroon, KR: Korea, IT: Italy, NE: Netherlands, FR: France, ES: Spain). Indian strains were indicated by ■ (Punjab) and ● (New Delhi).

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Fig. 3: Plausible route of spread of CRF02_AG strain in Africa, Europe and Asia

Note: The route of spread of 02_AG strain from West and Central West Africa to Europe (Netherland, France, Spain, Sweden) and Asia (Punjab and Delhi of India) is shown by arrows. (External boundaries of the countries shown in the map as depicted are neither correct nor authentic)

The most remarkable finding was the predominance of CRF02_AG strain among the unrelated Commercial Sex Workers (CSWs) from Delhi (Capitol of India) region. Most of the isolates from Punjab, Himachal Pradesh and Haryana region showed relatedness with consensus C. We also found one CRF02_AG (FJ769837) from Punjab (the rest from CSWs) and one subtype B (FJ769836) envelope-specific sequence from Himachal Pradesh.

The neighbor joining tree conducted in MEGA4 of 02_AG strains showed clustering of Indian CRF02_AG isolates with African CRF_02 clades (Fig. 2). Isolates from Punjab region formed a direct branching with two Nigerian isolates (accn. no. DQ168577 and DQ168578) with high bootstrap value (87, >70 considered significant) whereas isolates from Delhi formed a monophyletic cluster with the reference strains reported from Djibouti, Niger and Ghana. Therefore, it is tempting to speculate that Indian 02_AG strains may have spread from Africa. The UPGMA and ME methods of analysis also gave similar results (data not shown). Blast search also showed greater relatedness of Indian strains with African strains (~90-93%) compared to the European strains.
Fig. 4: Nucleotide distance (Kimura two-parameter model) comparisons within V3-V5 sequences of subtype C and Indian CRF02_AG strains

Note: Numbers within columns are the mean distances and error bars are standard errors of the mean. p-values refer to difference between group’s means (unpaired t-test with Welch’s correction). Mean and pairwise distances were calculated using the Poisson correction method in MEGA4

Taking into consideration the origin and movement of CRF_02AG in Europe and Asia, we predict the most likely route of introduction of this strain in India in Fig. 3. Since we found equal number of subtype C and CRF02_AG (6 each) sequences, we analyzed the intra-subtype differences between these strains. The analysis of intra-subtype nucleotide distances revealed that the subtype C population is significantly more divergent and diverse than CRF02_AG population (p<0.004; Fig. 4).

DISCUSSION

Although subtype C and other recombinant forms of different HIV-1 strains have been reported from different regions from India, this is the first report of CRF02_AG strain from India. This probably indicates a recent introduction of CRF02_AG strain in India. Co-circulations of variety of genetic subtypes were reported from different states of India from the very beginning of HIV-1 pandemic. We recently reported the presence of B/C recombinant LTR (Neogi et al., 2008) and B/C/D recombinant Vpr from north India (Bano et al., 2009). The geographical location of North India plays an important role in influencing the magnitude and nature of HIV-1 epidemic of India. This region is surrounded by China, Pakistan and Nepal where the HIV-1 epidemic is driven by different genetic subtypes (Pakistan: A and China: 07_BC and 08_BC). Therefore, due to extensive traveling, tourism and trade, by HIV-1 infected individuals of the neighboring countries, ideal situation may be created for generating novel and complex recombinant strains. The epicenter of global spread of 02_AG strain is West and central west of Africa. The Indian 02_AG strain has very likely spread from this region of Africa. This strain has potential to recombine with the most dominant subtype of India, the subtype C, along with the various recombinants that have been reported to date from India (Neogi et al., 2009). Our data also underscores the importance of rapidly evolving scenario of HIV-1 epidemic in this region which will have enormous impact on the design of vaccine antigens.

CONCLUSION

In conclusion, we show for the first time presence and transmission of HIV-1 02_AG strain in India and presence of subtypes B and C in North India. These observations will impact on the T-cell epitope based vaccine. The existence of multiple HIV-1 genetic subtypes in this region is likely to generate novel and complex recombinants. Since this is the first report of CRF02_AG from India, it will be important to monitor the spread of this strain and its impact on HIV-1 transmission in India.

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