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Nasal Myiasis Caused by Cochliomyia Hominivorax in the United States: A Case Report

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Abstract: Cases of human myiasis caused by *Cochliomyia hominivorax* are typically reported in Central and South America. We report a case of nasal myiasis, which manifested in a hospitalized patient in the United States who did not have a recent travel history. The larvae were discovered in the patient's nares on the fifth day of hospitalization. The patient was successfully treated with Ivermectin over two days.

Key words: Nasal myiasis, Cochliomyia hominivorax, screw-worm, ivermectin

INTRODUCTION

Myiasis is defined as the consumption of tissues, fluids, or ingested foods of vertebrates by the larvae of organisms of the order *Diptera* (Hall and Wall, 1995). While myiasis typically involves infestation of dead tissues, *Cochliomyia hominivorax*, the New World screw-worm fly, is an obligate parasite that preferentially consumes the living tissue of warm-blooded mammals. Typical infestations occur in livestock and manifest as weight loss (Madeira *et al.*, 1998).

Cases of *Cochliomyia hominivorax* infections in humans are less common and occur in patients with the following risk factors: open wounds, poor hygiene, low educational level, alcoholism, immobility and physical or mental disability (Batista-Da-Silva *et al.*, 2011).

Case: The patient is a 70 year-old-man with a past medical history of diabetes mellitus, hypertension, Chronic Obstructive Pulmonary Disease (COPD) and untreated Obstructive Sleep Apnea (OSA). He was admitted to the hospital for shortness of breath and intermittent hyper-somnolence. He was subsequently intubated due to hypercarbic respiratory failure and was transferred to the Intensive Care Unit.

On day five of hospitalization, a nurse observed white thread-like organisms emerging from the patient's nasal cavity and entering the patient's mouth Fig. 1. The same organisms were found in the suction tubing. Nasal swab was performed and a dozen larvae were collected and sent to microbiology. Upon visualization with direct light, the organisms displayed photophobia by migrating rapidly back into the nasal cavity. The patient's nares were flushed with mineral oil and he received Ivermectin 200 mcg kg⁻¹ per day for two days.



Fig. 1: Maggots visualized in patient's nasal cavity

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The following day, maxillofacial and head CT showed layering of fluid and mucosal thickening of the maxillary, ethmoid and sphenoid sinuses without evidence of deep tissue involvement. Augmentin 875mg twice a day was initiated.

Based on morphology, specimens sent to microbiology the previous night were confirmed to be *Cochliomyia hominivorax* Fig. 2 and 3. The case was reported to the hospital Infection Control Department and the State Department of Health. On hospital day 8, the patient was evaluated by fiberoptic nasal endoscopy, which revealed diffusely hyperemic nasal mucosa with old blood in the posterior nasopharynx, but without significant ulceration or necrosis. No larvae were seen.

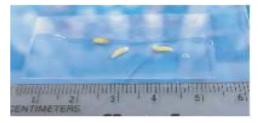
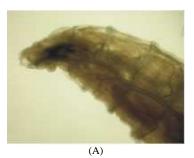


Fig. 2: Gross morphology of *Cochliomyia* hominivorax larvae



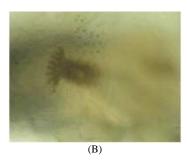


Fig. 3: (A) Microbiological analysis of *Cochliomyia* hominivorax under the microscope (400×).
Posterior spiracles and posterior surface of the last segment (B) Open peritreme of posterior spiracle Prior to its eradication from the United States in 1982, *Cochliomyia hominivorax* caused significant economic losses to the livestock industry (Wyss, 2000). Detection of *Cochliomyia hominivorax* on livestock in the United States is reportable to the State Veterinarian USDA Animal and Plant Health Inspection Service 2011. Since 1982, there have been several case reports of myiasis involving *Cochliomyia hominivorax* in imported animals from endemic countries (APHIS, 2007).

Cases of screw-worm myiasis in humans continue to be reported in endemic countries, especially in Central and South America (Powers *et al.*, 1996; Gomez *et al.*, 2003; Duque and Ardila, 2011; Coronado and Kowalski, 2009). Human myiasis has also been reported in individuals who have traveled to these countries (Mehr *et al.*, 1991). The larvae can cause ocular (Khurana *et al.*, 2010), oral (Junior *et al.*, 2010), umbilical (Duro *et al.*, 2007), subcutaneous (Trombetta *et al.*, 2009) and nasal (Couppie *et al.*, 2005) infections. The *Cochliomyia hominivorax* adult fly form lays eggs on the edges of wounds and necrotic tissue; the larval form consumes living tissue resulting in destruction of soft tissues, including cartilage (Hall and Wall, 1995; Gomez *et al.*, 2003).

The patient is originally from Puerto Rico and denied recent travel outside the United States. Friends from South America, who came in contact with the patient several days prior to hospitalization, are a possible source of the parasite. Typically, within 12-24 hours of the female fly depositing eggs, larvae emerge and feed on the host (IOEIC, 2008). After five to nine days, the larvae mature and abandon the host to pupate in soil (IOEIC, 2008). Manifestation of the larvae outside the patient's nares on the fifth day of hospitalization suggested that the infestation most likely occurred outside the hospital and the patient's altered mental status at home may have predisposed him to infestation.

Typically, myiasis is treated with direct removal of larvae, systemic treatment with Ivermectin 150-200 mcg kg⁻¹ over two days and topical adjuvant antibiotics (Rossi-Schneider *et al.*, 2007; Maheshwari and Naidu, 2010). Application of ether, chloroform, or oil may also be used to coax the larvae out of the subcutaneous tissue, (Maheshwari and Naidu, 2010) but the utility of this approach is unclear (Shinohara *et al.*, 2004). With nasal myiasis, endoscopy is often utilized to evaluate the nasal cavity and extract larvae under direct visualization (Tsang and Lee, 2009). Maxillofacial CT scans are often useful in nasal myiasis to determine the extent of soft tissue or bony destruction (Lee *et al.*, 2011).

To our knowledge, this is the first reported case of human nasal myiasis by *Cochliomyia hominivorax* in

the United States, since its eradication in 1982. Treatment with Ivermectin appears to have been effective as no larvae were found after 48 hours of treatment initiation and the patient remained without evidence of infection two weeks later.

With treatment of COPD and OSA, the patient improved and was discharged to a rehabilitation facility 21 days post admission.

CONCLUSION

With globalization and facillitated travel, remergence of previously eradicated organisms, such as *Cochliomyia hominivorax*, may be more likely to occur. We report a case of nasal myiasis caused by *Cochliomyia hominivorax*, which was evaluated with head and maxillofacial CT and nasal endoscopy and successfully treated with a two day course of Ivermectin.

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