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# Effect of Fig (*Ficus carica*) Leaf Extract on the Secretion and Content of Cholesterol in Hepg2 Cell

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Abstract: Traditional medicines remain a source of potential for discovering of new compounds with valuable pharmacological activities. Leaves of Ficus carica were dried, powdered and extracted using methanol (ME). An aliquot of ME was dried and re-extracted by water: chloroform and the other aliquot by water: peteruleum ether. Effect of aqueous fractions of the former (ACR; 0.08, 0.1 and 0.13 mg dL<sup>-1</sup>), the latter (APR; 0.07, 0.1 and 0.15 mg dL<sup>-1</sup>) and ME (0.03, 0.05 and 0.08 mg dL<sup>-1</sup>) of *Ficus carica* leaf on the secretion and cell content of cholesterol in HepG2 cells were studied. Extracts were added to the media in both basal and glucose stimulated conditions and incubated for 48h. While glucose significantly increased cholesterol secretion (17±0.76 mg dL<sup>-1</sup>) *vs* basal condition (6.91±0.66 mg dL<sup>-1</sup>), co-incubation with extracts reduced secretion of cholesterol in many concentrations of the stimulated condition. On the other hand, cholesterol content of HepG2 in glucose stimulated condition (2.73±0.39 mg dL<sup>-1</sup>) showed significant increase compared to the basal status (1.96±0.14 mg dL<sup>-1</sup>) (p<0.001). Moreover such decrease was shown in response to many concentrations of the extracts. These properties making the hydro-extracts of fig leaf a potentially safe intervention to modulate postprandial hyperlipidemia.

### Key words: Ficus carica, leaf extract, cholesterol

#### **INTRODUCTION**

Hypercholesterolemia is one of the major risk factors in development of coronary artery disease in recent years<sup>[2,6]</sup>. Nowadays; many non-prescribed treatments have been available for lowering the cholesterol. In this regard, traditional medicines are pursued as alternative drugs for treatment of hypercholesterolemia. So far, many traditional medicines have been investigated for their therapeutic effects both in humans and in experimental animals<sup>[2,12,13,20,21]</sup>. The hypocholestrolemic effect of citrus peel extract<sup>[2]</sup>, stems of Salvadora persica<sup>[8]</sup>, aqueous extract of *Retama raetem*<sup>[12]</sup>, aqueous extract of *Triticum repens*<sup>[13]</sup> and *Anka* (a fermented rice product of *monascus* sp.)<sup>[20]</sup> have been shown in Rats. The hypocholesterolemic effect of Allium sativum, in human has proved for many years ago<sup>[6]</sup>. In addition, several therapeutic effects have been shown for Ficus carica, different parts of such as hypoglycemia<sup>[18]</sup>, cancer suppressive<sup>[16]</sup>, anthelmintic<sup>[5]</sup>, hypotriglyceridemia<sup>[1,14]</sup> hypocholestrolemia<sup>[15]</sup> and bovine papilomatosis<sup>[10]</sup>.

Canal *et al*<sup>[4]</sup> showed that chloroform extract obtained from a decoction of *Ficus carica* leaves improved the blood cholesterol status in streptozotocin induced diabetic rats<sup>[4]</sup>. Furthermore, Shukla *et al.*,<sup>[19]</sup> showed that bark aqueous extract of *Ficus bengalensis* decreased serum low density lipoprotein cholesterol levels (LDL-C) (59%) and very low density lipoprotein cholesterol levels (60%) in hypercholesterolemic rabbits<sup>[19]</sup>. Thus, the aim of the present study was to extend the findings on the effects of *Ficus carica* leaf extract on the cholesterol status as secretion and content in both cell line and animal models and also to find its effective constituent.

### MATERIALS AND METHODS

The leaves of *Ficus carica* were collected from fig trees of Aminabad Institute, School of Veterinary Medicine, University of Tehran in July 2005 at four different times. All leaves were dried by air-flow in shadow, grinded and kept in a glass container. After then, five different extracts were prepared as follows: methanolic extract (ME) was prepared by using

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suxhelet apparatus model HP-6-500 and concentrated by rotary evaporator model Heidolph. The recent extract was re-extracted by choloroform (CRE) and peterullum ether (PRE), respectively. Again, each recent extract was re-extracted by distilled water and named as ACR and APR for choloroform and peterullum ether, respectively. All extracts were lyophilized by a lyophilizer (Epsilon-1-12D Chirst Company), rolled in aluminium foil and kept in -70°C until treatments.

Phytochemical analysis for alkaloid, saponin, tannin, flavonoid and glycoside was done<sup>[17]</sup>. HepG2 cells were prepared from Pasture Institute (Tehran, Iran) and cultured in T75 flasks containing DMEM+ 10% FBS + 1% L-glutamine. Incubation was done with concentration increasing of ME (0.03. $0.05, 0.08 \text{ mg dL}^{-1}$ , CRE (0.08, 0.01, 0.13 mg dL<sup>-1</sup>) and ACR  $(0.07, 0.1, 0.15 \text{ mg dL}^{-1})$ . Experiments were done both in basal and glucose stimulated conditions. Incubations were done for 48 hours at 5%CO<sup>2</sup> in a humid incubator. Then the cells and media were collected and their lipids content were extracted by Bligh and Dyer<sup>[3]</sup> method . Cholesterol levels were measured in the cell extract by the method of Fossati and Prencipe[1982]. Data were analyzed by 1-way ANOVA, for determination of difference between mean values using Sigma Stat Software. Alfa in all cases was 5% (p<0.05).

## RESULTS

Phytochemical analysis shows fig leaf extract (FTE) has minor, moderately and huge amounts of alkaloid, flavonoid and tannin, respectively. Effect of different levels of FTE on the secretion of cholesterol in HepG2 cells were shown in Table 1.

We showed ME (0.03, 0.05, 0.08 mg dL<sup>-1</sup>), ACR (0.08, 0.13 mg dL<sup>-1</sup>) and CRE (0.07, 0.15mg dL<sup>-1</sup>) increased cholesterol secretion higher than the basal levels (p<0.001).

Furthermore, effect of different levels of FTE on the secretion of cholesterol in stimulated condition was shown in Table 2. It can be seen that ME, APR and ACR can significantly decrease (p<0.001) secretion of stimulated cholesterol secretion to the basal levels.

Effect of different levels of FTE on the content of cholesterol in basal condition was shown in Table 2. We showed significant increase in the cholesterol content of the cell in response to ME (0.03 mg dL<sup>-1</sup>) (p<0.001) and ACR (0.07 mg dL<sup>-1</sup>) (p = 0.001). However, CRE decreased cholesterol values to below the basal levels (0.15 mg dL<sup>-1</sup>). On the other hand, CRE had no effect on the cellular cholesterol levels, when compared to the stimulated levels.

Table 1: Effect of different extracts of FTE on the secretion of		
cholesterol in both basal and stimulated conditions in		
HepG2 cells. Values were expressed as mean $\pm$ SD (n = 6).		
Different concentrations of each extract were compared		
with control by 1-way ANOVA		

	with control by I-way ANOVA				
		TG concentration	TG concentration		
		in basal	in stimulated		
		condition	condition		
Group	Status	(µg/well)	(µg/well)		
Ι	Control	6.91±0.66	17±0.76		
II	ME 0.03(%)	9.27±0.98	10.91±1.09		
III	ME 0.05(%)	10.36±0.65	7.64±0.53		
IV	ME 0.08(%)	7.64±0.49	6.55±0.6		
V	PRE 0.08(%)	8.73±0.37	6±0.54		
VI	PRE 0.1(%)	6.55±0.76	6.82±0.14		
VII	PRE 0.13(%)	7.64±0.7	8.18±0.71		
VIII	CRE 0.07(%)	6±0.42	10.09±0.6		
IX	CRE 0.1(%)	6.55±0.76	7.36±0.63		
Х	CRE 0.15(%)	8.73±0.65	6±1.14		
p values (differences among		I,II; I, III; I, V,	I, II; I,V; I, VII,		
different concentrations		I,VIII;I,X(<0.001),	I, VIII(0.001),		
of each extract)		I,IV; I, VII(0.013)	I, III(0.037),		
			I,X(0.007)		

Table 2: Effect of different extracts of FTE on the content of cholesterol in both basal and stimulated conditions in HepG2 cells. Values were expressed as mean±SD (n = 6). Different concentrations of each extract were compared with control by 1-way ANOVA

Group	Status	TG concentration	TG concentration
		in basal	in stimulated
		condition	condition
		(µg/well)	(µg/well)
Ι	Control	1.96±0.14	2.73±0.37
II	ME 0.03(%)	2.36±0.25	2.55±0.67
III	ME 0.05(%)	1.86±0.36	1.64±0.42
IV	ME 0.08(%)	1.73±0.21	2.18±0.4
V	PRE 0.08(%)	1.64±0.95	2.73±0.44
VI	PRE 0.1(%)	2.18±0.27	1.45±0.18
VII	PRE 0.13(%)	2.36±0.29	1.64±0.14
VIII	CRE 0.07(%)	2.55±0.49	2.91±0.35
IX	CRE 0.1(%)	2±0.6	1.64±0.23
Х	CRE 0.15(%)	$1.45 \pm 0.45$	1.45±0.39
p values (differences among		I,II; I,VIII(<0.001);	I,II; I,V; I,VI; I,
different concentrations		I,X(0.007)	VIII; I,X(<0.001);
of each extract)			I, VII(0.009);
			IX(0.022)

Effect of different levels of FTE extracts on the content of cholesterol in the stimulated condition was shown in Table 2. It can be seen that ME (0.05 and 0.08 mg dL<sup>-1</sup>) and ACR (0.1, 0.15 mg dL<sup>-1</sup>) reduced back the cellular cholesterol content to the basal levels.

#### DISCUSSION

In the most of experiments that have been done by different investigators on animal models, the levels of serum cholesterol have been increased in hypercholestrolemic or hyperlipidemic conditions<sup>[8,14,15,19]</sup> some studies on streptozotocin or

alloxan induced diabetic rats showed an increase in the levels of serum lipids, LDL-C and VLDL-C parameters<sup>[4,12,13]</sup>.On the other hand, some traditional medicines have effective components against to hyperlipidemic conditions. For example, aqueous extract of *Ratama reatem* decreased the levels of serum cholesterol in the streptozotocin induced diabetic rats<sup>[12]</sup>.Yang *et al.*,<sup>[21]</sup> showed that Paenoflorin which was isolated from the methanolic extract of *Paeonia lactifora* lowered serum cholesterol, LDL-C and trigelycerid levels in an experimentally induced hyperlipidaemic rats<sup>[21]</sup>. These investigations showed that some effective components in plants can decrease the lipid parameters in both hyperlipidemic and diabetic animals.

We showed that all aqueous extracts can significantly decrease (p<0.001) secretion of cholesterol from the liver cell in both stimulated and basal condition which is resemble to the diabetic animals. These findings are in good agreement with other findings<sup>[4,12,13]</sup>. However, Future studies will need to examine the mechanism of different FTE effects on the basal and glucose induced lipid changes to deduce if the effect is due to altered de novo cholesterol synthesis or increased catabolism of cholesterol.

Our phytochemical experiments showed that leaf of *Ficus carica* has huge amount of flavonoids. Lee *et al.*,<sup>[11]</sup> argued that naringenin 7-O-cetyl-synthetic drivate of naringenin is a flavonoid that can inhibit HMG-CoA reductase and ACAT activities in high cholesterol-fed rats<sup>[11]</sup>. Furthermore, it has been shown that tangerine peel extract and mixture of two citrus flavonoids (naringenin and hespridin) significantly lowered the levels of cholesterol in both plasma and liver. Hence, it can be deduced that inhibition of HMG-CoA reductase and ACAT activities may be contributed to the flavonoids in the fig leaf extract.

In conclusion these preliminary data suggest that hydroextract of fig leaf administration may be an alternative method to reduce hyperlipidemia, particularly postprandially induced ones.

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