

Phoretic Association and Facultative Parasitoidism Between *Megaselia scalaris* and Blowflies, Under Natural Conditions

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Abstract: Problem statement: This is the first time that a phoretic association and facultative parasitoidism between *Megaselia scalaris* and blowflies (Diptera: Calliphoridae) has been reported under natural conditions. **Approach:** All fly specimens were collected in the mangrove swamp in Itaboraí, RJ, Brazil. **Results:** A total of 291 flies were collected belonging to four (04) species of the family Calliphoridae: *Chrysomya megacephala* (Fabricius) (280), *Cochliomyia macellaria* (Fabricius) (05), *Chrysomya albiceps* (Wiedemann) (05), *Chrysomya putoria* (Wiedemann) (01) and all had third instar larvae of *M. scalaris* or had eggs of the Phoridae attached to their bodies. The specimens were separated and the flies, larvae and eggs were then identified. **Conclusion/Recommendations:** The highest prevalence for the capture of these flies was in the spring. From the study performed we conclude that *M. scalaris* acting as a facultative parasitoid of insects could be of great significance showing the need to screen laboratories more securely against invasions by *M. scalaris*.

Key words: APA guapi-mirim, diptera, mangrove swamps, reported finding, natural conditions

INTRODUCTION

Phoridae are small flies and have been described on exposed human corpses (Greenberg and Wells, 1998). About 225 genera and over 2,500 species of Phoridae are known in the world fauna (Borgmeier and Prado, 1975) and the genus *Megaselia* Rondani includes around 1,400 species distributed in the tropics and subtropics (Costa *et al.*, 2007).

Megaselia scalaris can be a parasitoid of insects of agronomic, veterinary and medical importance (Ulloa and Hernandez, 1981; Rocha *et al.*, 1984; Harrison and Gardner, 1991; Costa *et al.*, 2007). This study was carried out to report the occurrence of the scuttle fly, *M. scalaris* as a parasitoid of blowflies and also the occurrence of blowflies as phoretic of *M. scalaris* under natural conditions in Itaboraí, Brazil.

MATERIALS AND METHODS

All specimens were collected in a small part of the mangrove swamp in Guapi-Mirim Environmental Protection Area, in Itaboraí (RJ), which has a total area of 138.25 km² with 61.80 km² of mangrove swamps. Its geographical location is S 22° 39' 30" – 22° 46' 50" latitude and W 42° 57' 00" – 43° 06' 40"

longitude. To trap and collect the flies, four 35 cm high, 15 cm diameter plastic traps placed 100m apart were used. (trap models were according to Ferreira (1978) and modified according to Batista-Da-Silva *et al.*, 2010). The traps were suspended at a height of 1.20 m above the ground for a period of 48 h. and the bait used was based on fish (sardine) in decomposition (methods according to Batista-Da-Silva *et al.*, 2010). When setting up the traps, the bait was transported in closed containers thus avoiding any possible contamination.

At the collection time, all specimens were killed by asphyxiation with 70% ethanol inside the traps and all specimens were then put into plastic pots containing 70% ethanol for transportation to the Laboratory of Leishmaniasis Transmitters (Department of Medical and Forensic Entomology) at the Institute Oswaldo Cruz-IOC/FIOCRUZ, RJ, Brazil. At the laboratory the flies were separated and then identified using a stereoscopic microscope (80X) and the dichotomous keys for families and species according to Mello (2003); Disney and Ashmole (2004); Disney (2005); the eggs according to Greenberg and Kunich (2002); the larvae according to Sukontason *et al.* (2002). The flies parasitized were dissected and observed internally using a stereoscopic microscope.

Table 1: Total number and percentage of flies parasitized or with eggs of the *Megaselia scalaris* attached to their bodies and caught during the different seasons of the year, in a mangrove swamp, Itaboraí, Rio de Janeiro, Brazil

Parasitized species caught through the year in a mangrove swamp										
Species	Spring		Summer		Autumn		Winter		Total	
	N	(%)	N	(%)	N	(%)	N	(%)	N	(%)
<i>Chrysomya albiceps</i>	02	0.72	01	16.67	02	33.34	00	0.00	05	1.72
<i>Chrysomya megacephala</i>	272	97.49	04	66.66	04	66.66	00	0.00	280	96.22
<i>Chrysomya putoria</i>	00	0.00	01	16.67	00	0.00	00	0.00	01	0.34
<i>Cochliomyia macellaria</i>	05	1.79	00	0.00	00	0.00	00	0.00	05	1.72
Total	279	100.00	06	100.00	06	100.00	00	0.00	291	100.00

RESULTS

A total of 291 flies were captured belonging to four (04) species of the family Calliphoridae (Table 1): *Chrysomya megacephala* (Fabricius) (280), *Cochliomyia macellaria* (Fabricius) (05), *Chrysomya albiceps* (Wiedemann) (05), *Chrysomya putoria* (Wiedemann) (01) and all had third instar larvae of *M. scalaris* (Fig. 1) or had eggs of the Phoridae (Fig. 2) attached to their bodies. The highest prevalence for the capture of these flies was in the spring (279).

The parasitized flies had part of his muscles and internal organs destroyed. The mean temperature during the collection was 31.63±4.0°C and relative humidity was 64.53 ± 10.9%. All Phoridae adults found inside the traps were identified as *M. scalaris* and the species *C. megacephala* showed higher occurrence of the third instar larvae of *M. scalaris* or eggs of the Phoridae parasitoid attached to their bodies.

DISCUSSION

The species *M. scalaris* have a wide geographical distribution (Borgmeier, 1968; Costa *et al.*, 2007) and according to Manix (1964) can be found in the tropics and subtropics.

The occurrence of this species infesting and laying eggs, under natural conditions, on the indigenous screwworm, *C. macellaria* and blowfly *Chrysomya* species of the family Calliphoridae has not been reported until now. The occurrence of this infestation may be related to the substrate (sardine in decomposition) used to attract the muscoid, which may have also attracted the species *M. scalaris*, who have part of their life cycle in decomposed organic matter. All adult Phoridae found inside the trap belonged to the species *M. scalaris* demonstrating an absence of competition with other Phoridae species. The identification of the adult is one of the easiest and most secure ways to identify small flies (Disney and Munk, 2004) and this fact confirmed that the eggs and larvae found on the Calliphoridae, belonged to the species *M. scalaris*.

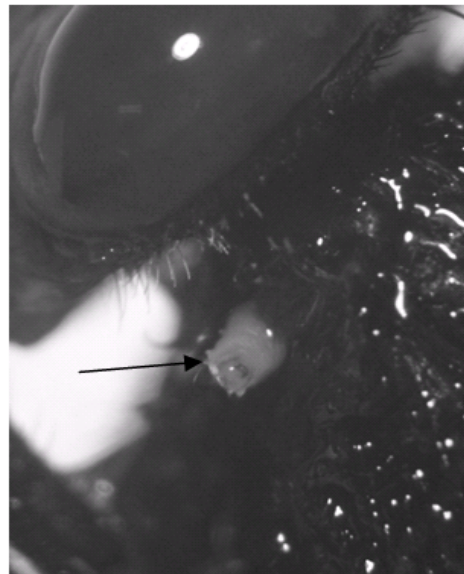


Fig. 1: *Chrysomya megacephala* with third instar larvae of *Megaselia scalaris*

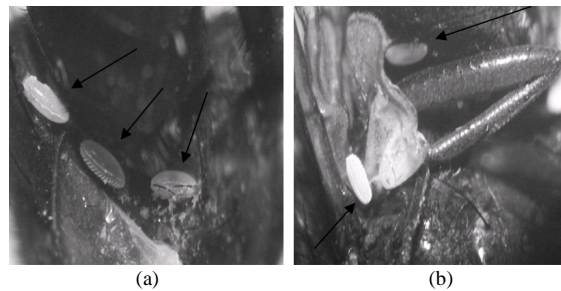


Fig. 2: *Chrysomya megacephala* (a) and *Chrysomya putoria* (b) with eggs of the parasitoid *Megaselia scalaris* attached to their bodies

According to Manix (1964) their life cycle is approximately 18 days for males and 20 days for females under 28°C (seven days in three larval instars). Although the traps used were exposed for 48 h periods, the larvae of *M. scalaris* had probably already

parasitized the family Calliphoridae flies before entering the traps. Spring and summer are the seasons with the highest tides in the mangrove swamp, with a maximum variation of 1.40 m, twice a day. The spring also represents the end of diapause for some species of the family Calliphoridae, which are well adapted to mangroves swamp. According to Greenberg and Kunich (2002) the pupae of this family can resist drowning for a period of four days. The abundance of Calliphoridae in mangrove swamps is higher in spring than in the other seasons. Coincidentally the period of highest occurrence of Calliphoridae acting as phoretic or suffering parasitoidism by *M. scalaris* larvae is also in the spring, which may indicate the synchronism of the life cycle of *M. scalaris* in the spring when the tides are highest. *M. scalaris* feeds on insects of many orders and families and included in the Coleoptera order are: Cicindellidae (Kirk-Spriggs and Marais, 1999), Scarabaeidae and Tenebrionidae (Batist, 2002). Also in the order Diptera *M. scalaris* feeds on Drosophilidae and Muscidae (Zwart *et al.*, 2005). Also *M. scalaris* has been associated with the laboratory cockroach (Dictyoptera) (Robinson, 1975; Miller, 1979), Gryllidae (Batist, 2002) and colonies of *Triatoma brasiliensis* (Costa *et al.*, 2007).

Megaselia scalaris infests cultures that are inadequately protected causing facultative parasitoidism (Disney, 2008).

CONCLUSION

The knowledge of the biological cycle of necrophagous entomofauna is one of the bases of forensic entomology and many of these insects are bred in laboratories and the occurrence of *M. scalaris*, as reported in this study could be of great significance showing the need to screen laboratories more securely against invasions by *M. scalaris*.

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