

# Co-Inhibitory Molecule Programmed Death-1 and its Ligands: A New Alternative Therapy for Human Immunodeficiency Virus Infection?

Leon-Flores A., P. Del Rio and G. Reyes-Teran

Instituto Nacional de Enfermedades Respiratorias,  
"Ismael Cosío Villegas", Mexico D.F., Mexico

Received 2012-05-31, Revised 2012-12-27; Accepted 2012-12-28

## ABSTRACT

Chronic viral infections are characterized by the up-regulation of a set of immunomodulatory receptors. The over-expression of co-inhibitory molecules on T cells leads to a dysfunctional T cell response with an "exhausted" phenotype. Programmed Death-1 (PD-1) is a molecule that exerts an inhibitory signal on the T cell receptor when it binds to the PD-L1 or PD-L2 ligands present on antigen-presenting cells. Also, the expression of these molecules has been associated to the loss of T cell functions as well as clinical markers of the progression of HIV infection. The study of these molecules has gained attention due to reports indicating that blockade of PD-1 pathway could partially reconstitute T cell functions. In fact, this mechanism has been proposed as an alternative treatment for some chronic viral infections such as HIV infection. This review is focused on those mechanisms that might be favouring the over-expression of PD-1 and its ligands during HIV infection and on the possible new approaches that, by reducing its expression, might represent new strategies for the treatment of HIV infection. Knowing the exact mechanism leading to PD-1, PD-L1 and PDL2 expression in physiologic and pathological conditions is essential for the development of successful treatments. Novel molecular mechanisms inhibiting PD-1 activation might have potential therapeutic use not only in HIV infection but also in other diseases.

**Keywords:** HIV, T-Cell Dysfunction, Programmed-Death 1 (PD-1), Programmed-Death Ligand (PD-L1), Human Leukocyte Antigen (HLA), Myeloid-Derived Suppressor Cells (MDSC)

## 1. INTRODUCTION

Despite extensive research on the Human Immunodeficiency Virus (HIV) for over 30 years, eradication of HIV-1 infection and treatment of AIDS remain a long-term challenge UNAIDS, 2010. The virus has developed strategies to spread and persist on infected tissues. On this basis, understanding the alterations to the immune system caused by the virus is one of the major goals for creating an effective treatment or vaccine. CD4<sup>+</sup> and CD8<sup>+</sup> T cells play an active role in the control of viral infections by producing soluble factors such as cytokines and chemokines and through the cytotoxic activity against infected cells mediated by

molecules such as granzyme and perforin (Freel *et al.*, 2011; Swain *et al.*, 2012). The T cell function against viruses and other microorganisms relies on the cellular activation status. According to the classic model, T cell activation results from the interaction of two signals (Baxter and Hodgkin, 2002): The first one ensures specificity by Antigen Presenting Cells (APC), which presents viral peptides processed and bonded to class Human Leukocyte Antigen (HLA) molecules to specific T Cell Receptor (TCR). However, this first signal does not trigger complete activation of T cells in absence of a second signal. Co-stimulatory molecules belonging to the B7/CD28 family are able to provide this second signal. Adequate stimulation of T cells through both

**Corresponding Author:** P. Del Rio, Instituto Nacional de Enfermedades Respiratorias, "Ismael Cosío Villegas", Mexico D.F., Mexico

signals leads to clonal expansion and promotes effector functions against pathogens. Both processes are accompanied by cell surface phenotype changes characteristic of cellular activation. In absence of a second signal T lymphocytes become anergic and their functions are limited (Wells, 2009). Co-stimulatory signals are classified as positive or negative depending on their effect on the TCR signalling. In this context, the co-stimulation process involves surface proteins that reduce or amplify the signal given by the TCR complex modulating T cell responses. One remarkable property of some members of this family is their capacity for providing co-inhibitory or co-stimulatory signals, depending on the binding site of the protein and on the type of receptor or ligand participating in the interaction (Alegre *et al.*, 2001; Carreno and Collins, 2002). Co-stimulatory signals are fundamental for physiological processes such as maintenance of immune tolerance (Bour-Jordan *et al.*, 2011), T-cell development (Keir *et al.*, 2005), pathogen-induced T cell activation (Paterson *et al.*, 2009) and immune response limitation once the antigen has been eliminated, preventing host damage (Iglesias-Chiesa *et al.*, 2008). In some situations, these molecules also contribute to pathological conditions such as the generation of autoimmunity and inflammation implicated in human rheumatoid arthritis (Raptopoulou *et al.*, 2010), atherosclerosis (Gotsman *et al.*, 2008), systemic lupus erythematosus (Hu *et al.*, 2004), multiple sclerosis (Wiendl *et al.*, 2003), parasitic infections (Bhadra *et al.*, 2012) and certain cancers (Lee *et al.*, 2010). In addition, an over expression of co-inhibitory proteins has been consistently documented in chronic viral infections (Jin *et al.*, 2011). As an example, HIV infection leads to a major expression of PD-1 and its ligands PD-L1 and PD-L2 (Yamamoto *et al.*, 2011), with important implications on the immune response (Yamamoto *et al.*, 2011). The over expression of PD-1 on CD4<sup>+</sup> and CD8<sup>+</sup> specific T cells during HIV infection is associated with a decrease in cytokine production, a gradual loss of CD8<sup>+</sup> T cell cytotoxic function and impaired cell proliferation, rendering T cells unable to control viral infection (Day *et al.*, 2006; Petrovas *et al.*, 2006; Trautmann *et al.*, 2006). In recent years, consensus has been reached on the relevant role of PD-1 as a negative regulator of T cell response during HIV infection; blockade of the intracellular pathway regulated by this molecule leads to restoration of certain T cell functions, representing novel approaches for the treatment of HIV infection (Porichis and Kaufmann, 2012) and for several other pathologies. There are few publications about mechanisms leading to the expression of PD-1 and its ligands; knowledge of

these mechanisms in physiologic and pathological conditions is essential for the development of successful treatments that, together with the antiretroviral treatment, would improve the immune response. This review is focused on the mechanisms that induce expression of PD-1 and its ligands in homeostatic processes and in pathological conditions. Novel molecular mechanisms inhibiting PD-1 activation with potential therapeutic use in different diseases are also discussed here.

### 1.1. Molecular Characteristics of PD-1 and its Ligands

The immunoreceptor Programmed Death-1 (PD-1; PDCD1; CD279) is a 50-55 kDa type 1 transmembrane glycoprotein of 288 amino acids (aa) and is a member of the immunoglobulin (Ig) super family. The PD-1 gene (*Pdcd1*) has an extension of 2106 cDNA nucleotides, is located in the human chromosome 2q37.3 (Shinohara *et al.*, 1994) and is encoded by 5 exons (Finger *et al.*, 1997; Ni *et al.*, 2007). PD-1 has a transmembrane region, an extra cellular region and a cytoplasmic region. The extracellular part of PD-1 is composed of approximately 147aa forming a single extracellular immunoglobulin with a variable-like domain (IgV-like) responsible for PD-1 binding with their specific ligands: PD-L1 (B7-H1; CD274) and PD-L2 (B7-DC; CD273) (Lazar-Molnar *et al.*, 2008; Zhang *et al.*, 2004). The transmembrane domain of PD-1 is composed of two loops of 27aa: The first is located in the middle of the protein and plays a role in anchoring the cell membrane; the second is located at the N terminus and is presumed to be a signal peptide (Zhang *et al.*, 2004). PD-1 cytoplasmic domain is composed of 94aa, responsible for triggering a signal through the activation of two tyrosine motifs: An ITIM (immunoreceptor tyrosine motif) and an ITSM (immunoreceptor tyrosine based switch motif) (Finger *et al.*, 1997; Ishida *et al.*, 1992). PD-1 has four potential post-transcriptional glycosylation sites in positions 49, 58 and 74, 116 (Finger *et al.*, 1997). PD-1 is considerably different from other B7/CD28 family members (e.g., CTLA-4 and ICOS), as exemplified by the lack of a proline-rich ligand recognition loop and the absence of a cysteine residue responsible for a disulfide bond formation (Zhang *et al.*, 2004). PD-1 is monomeric in solution and on cell surface, contrasting with other co-inhibitory molecules such as CTLA-4 and ICOS, which are presented as disulfide-linked homodimers (Mages *et al.*, 2000; Schwartz *et al.*, 2001; Zhang *et al.*, 2004). High-resolution crystallographic analysis revealed that PD-1/PD-L2 complex is different from CTLA-4/B7

inhibitory complexes in both, overall organization and particular molecular interactions, responsible for binding and specificity (Lazar-Molnar *et al.*, 2008).

The ligands of PD-1 are also well described, the type 1 transmembrane glycoprotein PD-L1 belongs to the immunoglobulin super family and is composed of 229 aa (Dong *et al.*, 1999). The PD-L1 gene (Cd274) is located in the chromosome 9p24.2 and is composed of seven exons (Ni *et al.*, 2007). This protein has an extracellular region of 122 aa, formed by two immunoglobulin-like domains: An IgC2-like domain and an IgV-like domain, with a potential transmembrane region of 21 hydrophobic aa and a small intracellular tail of 31 aa. PD-L1 has four potential glycosylation sites located in position 35, 192, 200 and 219 of the aa sequence with four disulfide bonds at position 40, 114, 155 and 209 (Dong *et al.*, 1999). There are some registered isoforms of PDL1, all of them produced by alternative splicing (He *et al.*, 2005). PD-L2, the second known PD-1 ligand, is a type 1 glycoprotein of 273 aa belonging to the immunoglobulin super family; the topographic protein sequence is constituted by an extracellular immunoglobulin with an Ig-like C2 type and a Ig-like V type domains, that contain a transmembrane region of 21 aa and an intracellular tail of 32 aa (Lazar-Molnar *et al.*, 2008). PD-L2 has some potential glycosylation sites at the 36, 64, 157, 163 and 189 positions, with four potential disulfide bonds at positions 42, 102, 143 and 192, with protein isoforms (Lazar-Molnar *et al.*, 2008; Ni *et al.*, 2007). The PD-L2 gene (*Pdcd1lg2*) is located in the chromosome 9 comprising six exons close to PD-L1 gene by only 42kb (Lazar-Molnar *et al.*, 2008).

## 1.2. Expression of PD-1 and its Ligands

Upon activation, PD-1 is expressed on CD4+ and CD8+ T lymphocytes (Bennett *et al.*, 2003; Vibhakar *et al.*, 1997), on B cells (Agata *et al.*, 1996), on stimulated monocytes/macrophages (Huang *et al.*, 2009) and on NKT cells (Moll *et al.*, 2009). It can also be expressed on non-lymphoid organs such as retina (Chen *et al.*, 2009). PD-L1 has a wider range of expression including dendritic cells (Wolfe *et al.*, 2011), macrophages (Wagner *et al.*, 2010), neutrophils (Bankey *et al.*, 2010) and activated T and B-lymphocytes (Francisco *et al.*, 2010; Rosignoli *et al.*, 2009). PD-L1 can also be expressed on pancreatic islet cells (Rajasalu *et al.*, 2010), keratinocytes (Youngnak-Piboonratanakit *et al.*, 2004), vascular endothelial cells (Mazanet and Hughes, 2002), astrocytes (Lipp *et al.*, 2007), cardiac endothelial cells (Grabie *et al.*, 2007), placenta (Petroff and Perchellet,

2010) and kidney (Menke *et al.*, 2007). The diversity of cells expressing PD-L1 suggests that it could have a role not only in APC but also in cells closely related to the immune system. PD-L2 is expressed in low levels on antigenpresenting cells (Tseng *et al.*, 2001), bone marrow-derived mast cells (Nakae *et al.*, 2006) and on activated human T cells (Ishida *et al.*, 2002; Messal *et al.*, 2011). Despite the well-defined expression of PD-1 and its ligands, little is known about the mechanisms regulating its expression. It is known that PD-1 can be induced with T-cell activation and by anti- and pro-inflammatory cytokines such as INF $\alpha$ , IL-10 and gamma chain cytokines IL2, IL-7, IL15, IL21 (Kinter *et al.*, 2008). Additionally, epigenetic mechanisms are involved in the control of PD-1 (Zhang *et al.*, 2011). It was recently shown that DNA methylation down regulates PD-1 expression resulting from viral-induced TCR activation; as the virus is cleared and TCR stimulation decreases, the PDCD1 gene is remethylated and the expression of PD-1 is down regulated (Youngblood *et al.*, 2011). Sustained demethylation of PDCD gene was observed in T cells during chronic virus infections and, consequently, increased PD-1 expression was also observed. PD-1 and PD-L1 are also up regulated on APC and CD8+ T cells after stimulation with TLR ligands (Berthon *et al.*, 2010; Meier *et al.*, 2008; Wong *et al.*, 2009). PD-L2 is mainly regulated by cytokines such as IL-10, IL4 and INF- $\gamma$  (Rodriguez-Garcia *et al.*, 2011; Stanciu *et al.*, 2006). Apparently, IFN- $\gamma$  is also one of the main regulators of PD-L1 expression in a wide range of cell types (Berthon *et al.*, 2010; Chen *et al.*, 2012; Gong *et al.*, 2009; Liang *et al.*, 2009). Further investigation on the mechanisms of PD-1, PD-L1 and PD-L2 expression and regulation is required to understand the role of these molecules in pathological processes.

## 1.3. PD-1 Activation by its Ligands

The effect of PD-1 binding to its ligands has been well characterized in T cells and B cells during chronic viral infections. PD-1 activation has been associated with T cell dysfunction due to inhibition of the TCR signalling and to altered expression of proteins and genes. As mentioned before, there are two main ligands for PD-1, being PD-L2 the one with more affinity for PD-1 than PD-L1 (Youngnak *et al.*, 2003). After binding with its ligands, PD-1 activation transmits an inhibitory signal to the TCR or BCR signal and down-modulates TRC complex (Karwacz *et al.*, 2012; Latchman *et al.*, 2001; Okazaki *et al.*, 2001; Pentcheva-Hoang *et al.*, 2007). PD-1 ligation results in phosphorylation of the tyrosines located in its cytoplasmic domain and

recruitment of the SHP- 1 and SHP-2 to the C-terminal tyrosine in the ITSM (Chemnitz *et al.*, 2004). SHP-2 then dephosphorylates TCR-associated CD-3  $\xi$ , ZAP70 Kinase Ch (PKCh) and Erk (Sheppard *et al.*, 2004), resulting in downstream direct inhibition of PI3K signalling (Parry *et al.*, 2005; Riley, 2009). Triggering of PD-1 pathway leads to alterations on cell cycle progression (Latchman *et al.*, 2001), on cytokine production (Cho *et al.*, 2009; Latchman *et al.*, 2001) and cells become susceptible to death by apoptotic mechanisms (Petrovas *et al.*, 2006). Specifically, blocking PI3K after PD-1 activation affects IL-2 production and glucose metabolism, compromising cell survival (Parry *et al.*, 2005). PD-1 pathway also prevents production of transcriptional factors associated with T cell effector functions such as GATA-3, T-bet and Eomes (Nurieva *et al.*, 2006). PD-1 activation induces a particular pattern of proteins and activation of genes. Studies on CD4+ T cells comparing the transcriptional profile during PD-1 inhibition *versus* other co-inhibitory transcriptional profiles such as CTLA-4 showed that PD-1 has a major influence in T cell activation (Parry *et al.*, 2005; Riley, 2009). The number of transcripts derived from the interaction of CD3 and CD28 is reduced in 67 and 90% by CTLA-4 and PD-1 respectively (Parry *et al.*, 2005; Youngblood *et al.*, 2012). CTLA-4 induces the expression of the anti-apoptotic gene Bcl-xL, while PD-1 does not. This might be the reason why PD-1 pathway renders T cells susceptible to apoptotic stimulus (Parry *et al.*, 2005).

#### 1.4. PD-1 and its Ligands During Chronic Viral Infections Such as HIV Infection

T cell exhaustion was initially observed in mice with LCMV infection (Zajac *et al.*, 1998). It is characteristic of an antigen-persistence environment caused by chronic infection with microorganisms such as, Epstein Barr Virus (EBV), Hepatitis B or C Virus (HBV or HCV), mycobacterium tuberculosis or HIV (Jin *et al.*, 2011). This phenotype of T cells is often called “exhausted” and is characterized by a progressive loss of T cell functions and a deteriorated immunological control of pathogens (Freeman *et al.*, 2006). The loss of functions is hierarchical, being the cytotoxic function, the proliferative capacity and IL-2 production lost at first, followed by the loss of TNF- $\alpha$  production and subsequently by the loss of IFN- $\gamma$  production (Wherry and Ahmed, 2004). Several extrinsic and intrinsic mechanisms are involved in the negative regulation of immune responses during an environment of antigen

persistence (Virgin *et al.*, 2009). Extrinsic mechanisms, referred as factors that extracellularly promote immune impairment through cell-cell or cell-protein interaction, include: (1) dysfunctional or suppressive Antigen-Presenting Cells (APCs) (Jin *et al.*, 2011; Rodrigue-Gervais *et al.*, 2010), (2) increase of regulatory cell populations just as regulatory T cells (Tregs) (Dolganic *et al.*, 2008; Manches *et al.*, 2008) and Myeloid-Derived Suppressor Cells (MDSC) (Chou *et al.*, 2012). Intrinsic mechanisms are referred as intracellular mechanisms that promote cell dysfunction, like the up-regulation of inhibitory receptors (Blackburn *et al.*, 2009; Fourcade *et al.*, 2010) and transcriptional repressor proteins (Shin *et al.*, 2009) due to an increased duration and magnitude of antigen stimulation, or by an environment enriched with immunosuppressive cytokines (Coussens and Werb, 2002; Virgin *et al.*, 2009). Thus, it has been suggested that the over-expression of co-inhibitory proteins is related to the dysfunction of T cells during chronic infections (Boni *et al.*, 2007; Day *et al.*, 2006; Freeman *et al.*, 2006; Petrovas *et al.*, 2006; Radziejewicz *et al.*, 2007; Trautmann *et al.*, 2006; Urbani *et al.*, 2006). Based on studies made in animal models and in humans, it is currently known that several chronic infections such as HCV and HBV up-regulate the expression of inhibitory proteins just as PD-1 and PD-L1 on virus-specific T cells (Trautmann *et al.*, 2006; Urbani *et al.*, 2006).

The effect of this protein up-regulation on T cell exhaustion has been extensively studied on HIV infection, becoming a potential hallmark of AIDS progression (Day *et al.*, 2006; Holm *et al.*, 2008). PD-1 overexpression on CD8+ and CD4+ T cells was first described by Petrovas *et al.* (2006), who showed that PD-1 favours the unresponsiveness of T cells against HIV. Since then, studies have focused on the patterns of expression and the relation of this protein with the functional status of T cells and on the screening of cells that express PD-1 and its ligands in several cohorts of HIV individuals (Barber *et al.*, 2006; Day *et al.*, 2006; Petrovas *et al.*, 2006; Porichis and Kaufmann, 2011; Trautmann *et al.*, 2006). PD-1 expression is dynamically regulated during T cell differentiation; from PD-1- naïve T cells to PD-1low memory T cells and PD-1hi effector T cells (Rosignoli *et al.*, 2009). PD-L1 is highly expressed on terminally differentiated effector T cells from viremic and aviremic individuals, while it is expressed in low levels by their naïve CD4+ T cells (Rosignoli *et al.*, 2009). PD-1 expression is higher in anatomic compartments with elevated viral replication such as lymph nodes, gut and peripheral lymphoid

tissues, than in peripheral blood cells (D'Souza *et al.*, 2007; He *et al.*, 2004; Velu *et al.*, 2009). However, it has not been defined if the higher expression of PD-1 in these anatomical compartments is also related to a more "exhausted" phenotype. The influence of the over-expression of these molecules on T cell functions has been clarified by using specific antibodies to block the interaction of PD-1 with its ligands (Finnefrock *et al.*, 2009; Freeman *et al.*, 2006). Blockade of the PD-1 pathway shows improvement in some T cell functions, enhances proliferation of HIV-specific CD4+ and CD8+ T cells (Porichis *et al.*, 2011) and increases the secretion of some cytokines (Muthumani *et al.*, 2011). Additionally, PD-1 expressed on monocytes might also be contributing to IL-10 production, which in turn favours T cell dysfunction (Said *et al.*, 2010). The effect of PD-1 on cellular cytotoxicity remains to be explored. One of the most relevant studies regarding blockade of PD-1 pathway is the one of Velu group in a model of SIV infection. This study showed an improvement of cellular and humoral responses not only in blood but also in gut (Velu *et al.*, 2009). Therefore, there is no doubt that the PD-1 pathway contributes significantly to the T cell dysfunction observed during HIV infection however, due to the increasing reports showing not only the increase of PD-1 expression on immune cells but also of its ligand PDL1, it is important to evaluate the role of its ligand in the context of HIV infection. Also, it might also be important to know other effects of the PD-1 activation pathway in order to revert T cell dysfunction. As an example, it is now known that PD-1 up-regulates a unique set of genes on exhausted T cells which includes BATF. Over-expression of BATF results in an exhaustion-like T cell phenotype, while silencing of BATF gene is able to restore the function of impaired T cells (Quigley *et al.*, 2010).

### 1.5. Factors Contributing to PD-1 and PD-L1 Over-Expression during HIV Infection

Although PD-1 expression has been described on cells of HIV+ individuals, little is known about the factors inducing PD-1 expression on HIV infection. Data indicate that the virus is the main factor inducing the expression of PD-1 and its ligands. HIV-specific CD8+ T cells express higher levels of PD-1 than HCV-, HBV or CMV- specific CD8+ T cells (Day *et al.*, 2006; Trautmann *et al.*, 2006), being the cells with immunodominant epitopes the ones that express higher levels of PD-1 (Conrad *et al.*, 2011). This might imply that HIV provides additional factors for the up-regulation

of PD-1 and possibly for its ligands. The expression of PD-1 on CD4+ and CD8+ T cells has a positive correlation with HIV plasma viral load and a negative correlation with peripheral blood CD4+ T cell count (Zhang *et al.*, 2007). PD-1 and PD-L1 are highly expressed on virus specific effector memory CD4+ T cells from acute and chronic infected subjects, while all T cell subsets of elite controllers (a minority group that naturally controls HIV replication) and uninfected subjects express PD-1 and PD-L1 in low levels (Porichis *et al.*, 2011; Rosignoli *et al.*, 2009), corroborating the idea that the virus directly influence the expression of such molecules. Correlation of PD-1 expression with viral load has been confirmed by studies describing that the reduced HIV-replication observed as a consequence of long-term combined Antiretroviral Therapy (cART), apparently decreases PD-1 expression on specific CD4+ T cells, leading to an increase in the number of functional T cells. Nevertheless, the fact that PD-L1 expression remains elevated on CD8+ T cells from viremic and aviremic subjects receiving cART (Rosignoli *et al.*, 2007), suggests a different mechanism of regulation for PD-L1 expression on these subjects. Furthermore, it has been proposed that viral load affects PD-1 expression in a context favouring chronic activation, a predominant feature of HIV infection (Estes *et al.*, 2008; Sauce *et al.*, 2007; Venkatachari *et al.*, 2008). The cell chronic activation characteristic of HIV infection results from the interaction of multiple factors like persistent antigenic load, cytokine environment and bacterial translocated products from gut or colorectal tissue in blood (Jin *et al.*, 2011). The close relation between activation markers and PD-1 over-expression suggests that chronic activation might also influence PD-1 expression during HIV infection (Hokey *et al.*, 2008; Holm *et al.*, 2008; Sauce *et al.*, 2007; Tendeiro *et al.*, 2012). On the contrary, other reports have shown that PD-1 is responsible of some characteristics of the hyper-immune activation (Holm *et al.*, 2008; Nakanjako *et al.*, 2011; Porichis *et al.*, 2011). Blockade of the PD-1 pathway during SIV infection reduces the expression of activation markers in T cells and, importantly, reduces the translocation of bacterial products in the colorectal tissue of rhesus macaques (Shetty *et al.*, 2012).

Although viral presence seems to be close related to PD-1 expression (Porichis *et al.*, 2011), there is no evidence that HIV-infection *per se* promotes PD-1 expression. The impact of HIV infection in PD-1 and PD-L1 has not been fully evaluated. In 2008 the group of Muthumani *et al.* (2008) reported that the viral protein

Nef induces the expression of PD-1 in CD4<sup>+</sup> T cells, representing a viral mechanism for inducing the expression of this protein. In contrast, the group of Venkatachari *et al.* (2008) reported in the same year that HIV infection down-regulates the expression of PD-1 in CD4<sup>+</sup> T cells, inhibiting apoptotic cell death. Differences in the expression of PD-1 depending on the type of virus, being the subtype D the one inducing higher expression of PD-1 in T cells than the subtype A, were subsequently reported (Bousheri *et al.*, 2009). The effect of HIV infection *per se* on PD-1 ligand expression is less clear; most studies have analysed the expression of these ligands in antigen presenting cells. It has been found that PD-L1 and PD-L2 can be induced in monocytes/macrophages after exposure to competent and inactivated HIV (Rodriguez-Garcia *et al.*, 2011). PDL1 could be preferentially up-regulated in macrophages in an anti-inflammatory environment and PD-L2 might be predominantly up-regulated in a pro-inflammatory environment (Rodriguez-Garcia *et al.*, 2011). HIV infection could also induce the expression of PD-L1 in DC (Muthumani *et al.*, 2011). Additionally, PD-L1 over-expression has also been observed in T cell subsets of HIV infected individuals (Rosignoli *et al.*, 2009), the mechanism by which this over-expression occurs remains poorly studied but, the over-expression of PD-1 ligands on T cells opens the possibility of a new T cell-T cell interaction that might be contributing to PD-1 up-regulation. Recently it has been shown that cytotoxic CD8<sup>+</sup> T cells obtain the PD-L1 molecule from mature DC by a new antigen specific mechanism called trogocytosis. Functional studies showed that the CD8<sup>+</sup> T cells that acquire PD-L1 from mature DC are able to induce apoptosis of CD8<sup>+</sup>PD-1<sup>+</sup> T cells (Gary *et al.*, 2011), opening the possibility for mechanisms based on cellular interactions for PD-L1 expression.

### 1.6. Inhibition of PD-1 Pathway as Alternative Therapy to Control HIV

Understanding the mechanisms responsible of the T cell dysfunction observed during chronic viral infections could provide novel therapeutic targets for the treatment of persisting infections. Accumulating evidence on the effect of PD-1 on T cells has increased the knowledge about the mechanisms of T cell exhaustion characteristic of chronic infections, placing PD-1 pathway as a novel strategy for HIV therapy (Porichis and Kaufmann, 2012). Several strategies are being developed for supporting the immune system not only in chronic viral infections but in other diseases affecting the immune system, some of them are: (1) blockade of PD-1/PD-L1 interaction with

specific antibodies or through other molecular approaches (Berger *et al.*, 2008; Brahmer *et al.*, 2010; Petrovas *et al.*, 2006); (2) use of PD-1 blockade as an adjuvant in ART treatment or in vaccines trials (Finnefrock *et al.*, 2009; Ha *et al.*, 2008); (3) blockade of several inhibitory receptors expressed on T-specific exhausted cells (Blackburn *et al.*, 2009; Jin *et al.*, 2010; Jones *et al.*, 2008); and (4) use of siRNA for PD-L1 in APC (Muthumani *et al.*, 2011).

The use of antibodies to prevent interaction of PD-1 with its ligands represents a novel strategy for T cell therapy, leading to restoration of T cell functions in a fraction of CD4<sup>+</sup> and CD8<sup>+</sup> T cells, improvement of cellular proliferation (Trautmann *et al.*, 2006) and a modest increase on cytokine production (Barber *et al.*, 2006; Petrovas *et al.*, 2006). However it has been suggested that inhibiting the PD-1/PD-L1 pathway may lead to sustained TCR signalling leading to hyper-activated T cells, or may generate inflammatory responses. Blockade of PD-1 pathway in tumour-bearing mice increases the cytotoxic responses against tumoral cells without remarkable autoimmune responses (He *et al.*, 2004). The clinical relevance of blocking this pathway is supported by pharmacokinetic studies in patients with hematologic malignancies (Berger *et al.*, 2008) or with solid tumours (Brahmer *et al.*, 2010). The effects of blocking PD-1 pathway in presence of chronic viral infections have not been assessed and the SIV infection model might be suitable for initiation of such studies. Additionally, the use of PD-1 blockade together with cART or vaccination may potentiate HIV-therapy as the presence of antigen would be reduced and the immune response would be stimulated (Finnefrock *et al.*, 2009). It has been shown that blockade of the PD-1 pathway in combination with vaccination synergistically enhance specific T cell response during LCMV infection and has an important impact on viral control (Ha *et al.*, 2008). Additional studies are required to assess the synergic effect of PD-1 blockade after cART initiation. It will be also interesting to know the effect of this blockade in T cells of other anatomic compartments in which antiretroviral treatment is poorly accessible, as well as to determine optimal timing for cART initiation and PD-1 blockade. The use of siRNA to avoid PD-L1 expression on APC has also been described as an alternative way to block PD-1 pathway and reverse T cell dysfunction (Borkner *et al.*, 2010; Iwamura *et al.*, 2012). Blocking PD-1 pathway, either through anti-PDL1 antibody or siRNA seems to be a good strategy to restore T cell functionality. However, PD-L1 also supplies a co-stimulatory signal by interacting with B7-1 and

considerations about silencing completely this molecule should also be taken. Further studies will elucidate possible alterations in this co-stimulatory signal caused by PD-L1 blockade.

Exhausted T cells express several co-inhibitory receptors showing that PD-1 is not the only molecule affecting T cell function. It is said that the more exhausted the cell is, the more co-inhibitory molecules are expressed, this molecules co-expression correlates with a faster progression to AIDS (Kassu *et al.*, 2010). Phenotype and gene expression profile studies showed the co-expression of several inhibitory receptors on exhausted CD4<sup>+</sup> and CD8<sup>+</sup> T cells, like T cell immunoglobulin mucin 3 (ITIM-3), Cytotoxic T-Lymphocyte Associated protein 4 (CTLA-4), natural killer cell receptor (2B4), Lymphocyte Activation Gene 3 (LAG-3), CD160, Leukocyte Immunoglobulin-Like Receptor superfamily B member 3 (LILRB3) and 4 (LILRB4), that act synergistically with PD-1 (Porichis *et al.*, 2011; Yamamoto *et al.*, 2011; Jones *et al.*, 2008). Thus, blocking more than one of these co-inhibitory receptors should improve not only the proliferative capacity of T cells (Yamamoto *et al.*, 2011) but also cytokine production and cytotoxic functions, strategy that has been less evaluated.

### 1.7. Final Remarks

In the last years, the PD-1 pathway has become a novel target for restoring the progressive T cell dysfunction observed on viral chronic infections. Combined antiretroviral therapy can achieve undetectable viral loads, but it is unable to eliminate HIV reservoirs or fully restore the immune system. Recent reports have shown that blocking PD-1 pathway can partially restore T cell functions. Because PD-1 has distinct roles in both physiological and pathological conditions, a better understanding of the molecular behaviour and biological functions of PD-1 and its ligands may contribute to the development of effective alternative or co-adjuvant therapies for HIV. As an example, there is a lack of knowledge on the cellular and molecular mechanisms affecting the cytotoxic function of CD8<sup>+</sup> T cells after activation of PD-1 intracellular pathways, an essential function for the elimination of infected cells. To our knowledge, only the group of Trautmann *et al.* (2006) has been able to show that blockade of PD-1/PD-L1 interaction slightly improves CD8<sup>+</sup> T cell degranulation. Additional studies exploring the expression of PD-1 and its ligands and their role in modulating the cytotoxic function of CD8<sup>+</sup> T cells are required. The mechanism by which PD-1 activation

modulates cytokine and other anti-viral factors production is also unknown. The fact that a dysfunctional cell not only expresses one inhibitory molecule but also a set of diverse inhibitory molecules emphasises the relevance of defining the T cell functions affected by the different phenotypic profiles. Exploring the clinical relevance of these events is essential for the knowledge of new targets and development of therapeutic models. PD-1 expression on HIV infection has been widely studied, but little is known about the expression and function of PD-1 ligands. The fact that a soluble form of PD-L1 has been found in plasma from patients with cancer (Chen *et al.*, 2012) represents a new mechanism of activating PD-1 pathway that may favours T cell dysfunction and mediate apoptotic mechanisms. Further studies are needed to elucidate PD-L1 and PD-L2 roles in HIV infection.

## 2. ACKNOWLEDGMENT

The researchers thank Claudia Alvarado de la Barrera for manuscript style correction. Authors thank Comisión de Equidad y Género de la H. Cámara de Diputados of México and Fundación México Vivo (<http://www.mexicovivo.org/>).

## 3. REFERENCES

- Agata, Y., A. Kawasaki, H. Nishimura, Y. Ishida and T. Tsubata *et al.*, 1996. Expression of the PD-1 antigen on the surface of stimulated mouse T and B lymphocytes. *Int. Immunol.*, 8: 765-772. DOI: 10.1093/intimm/8.5.765
- Alegre, M.L., K.A. Frauwirth and C.B. Thompson, 2001. T-cell regulation by CD28 and CTLA-4. *Nat. Rev. Immunol.*, 1: 220-228. DOI: 10.1038/35105024
- Bankey, P.E., S. Banerjee, A. Zucchiatti, M. De and R.W. Sleem *et al.*, 2010. Cytokine induced expression of programmed death ligands in human neutrophils. *Immunol. Lett.*, 129: 100-107. DOI: 10.1016/j.imlet.2010.01.006
- Barber, D.L., E.J. Wherry, D. Masopust, B. Zhu and J.P. Allison *et al.*, 2006. Restoring function in exhausted CD8 T cells during chronic viral infection. *Nature*, 439: 682-687. DOI: 10.1038/nature04444
- Baxter, A.G. and P.D. Hodgkin, 2002. Activation rules: The two-signal theories of immune activation. *Nat. Rev. Immunol.*, 2: 439-446. DOI: 10.1038/nri823

- Bennett, F., D. Luxenberg, V. Ling, I.M. Wang and K. Marquette *et al.*, 2003. Program death-1 engagement upon TCR activation has distinct effects on costimulation and cytokine-driven proliferation: Attenuation of ICOS, IL-4 and IL-21, but not CD28, IL-7 and IL-15 responses. *J. Immunol.*, 170: 711-718. PMID: 12517932
- Berger, R., R. Rotem-Yehudar, G. Slama, S. Landes and A. Kneller *et al.*, 2008. Phase I safety and pharmacokinetic study of CT-011, a humanized antibody interacting with PD-1, in patients with advanced hematologic malignancies. *Clin. Cancer Res.*, 14: 3044-3051. DOI: 10.1158/1078-0432.CCR-07-4079
- Berthon, C., V. Driss, J. Liu, K. Kuranda and X. Leleu *et al.*, 2010. In acute myeloid leukemia, B7-H1 (PD-L1) protection of blasts from cytotoxic T cells is induced by TLR ligands and interferon-gamma and can be reversed using MEK inhibitors. *Cancer Immunol. Immunother.*, 59: 1839-1849. DOI: 10.1007/s00262-010-0909-y
- Bhadra, R., J.P. Gigley and I.A. Khan, 2012. PD-1-mediated attrition of polyfunctional memory CD8+ T cells in chronic toxoplasma infection. *J. Infect. Dis.*, 206: 125-134. DOI: 10.1093/infdis/jis304
- Blackburn, S.D., H. Shin, W.N. Haining, T. Zou and C.J. Workman *et al.*, 2009. Coregulation of CD8+ T cell exhaustion by multiple inhibitory receptors during chronic viral infection. *Nat. Immunol.*, 10: 29-37. DOI: 10.1038/ni.1679
- Boni, C., P. Fisicaro, C. Valdatta, B. Amadei and P.D. Vincenzo *et al.*, 2007. Characterization of Hepatitis B Virus (HBV)-specific T-cell dysfunction in chronic HBV infection. *J. Virol.*, 81: 4215-4225. DOI: 10.1128/JVI.02844-06
- Borkner, L., A. Kaiser, W.V.D. Kastele, R. Andreesen and A. Mackensen *et al.*, 2010. RNA interference targeting programmed death receptor-1 improves immune functions of tumor-specific T cells. *Cancer Immunol. Immunother.*, 59: 1173-1183. DOI: 10.1007/s00262-010-0842-0
- Bour-Jordan, H., J.H. Esensten, M. Martinez-Llordella, C. Penaranda and M. Stumpf *et al.*, 2011. Intrinsic and extrinsic control of peripheral T-cell tolerance by costimulatory molecules of the CD28/B7 family. *Immunol. Rev.*, 241: 180-205. DOI: 10.1111/j.1600-065X.2011.01011.x
- Bousheri, S., C. Burke, I. Ssewanyana, R. Harrigan and J. Martin *et al.*, 2009. Infection with different hiv subtypes is associated with CD4 activation-associated dysfunction and apoptosis. *J. Acquir. Immune Defic. Syndr.*, 52: 548-552. DOI: 10.1097/QAI.0b013e3181c1d456
- Brahmer, J.R., C.G. Drake, I. Wollner, J.D. Powderly and J. Picus *et al.*, 2010. Phase I study of single-agent anti-programmed death-1 (MDX-1106) in refractory solid tumors: Safety, clinical activity, pharmacodynamics and immunologic correlates. *J. Clin. Oncol.*, 28: 3167-3175. DOI: 10.1200/JCO.2009.26.7609
- Carreno, B.M. and M. Collins, 2002. The B7 family of ligands and its receptors: New pathways for costimulation and inhibition of immune responses. *Annu Rev. Immunol.*, 20: 29-53. DOI: 10.1146/annurev.immunol.20.091101.091806
- Chemnitz, J.M., R.V. Parry, K.E. Nichols, C.H. June and J.L. Riley, 2004. SHP-1 and SHP-2 associate with immunoreceptor tyrosine-based switch motif of programmed death 1 upon primary human T cell stimulation, but only receptor ligation prevents T cell activation. *J. Immunol.*, 173: 945-954. PMID: 15240681
- Chen, J., Y. Feng, L. Lu, H. Wang and L. Dai *et al.*, 2012. Interferon- $\gamma$ -induced PD-L1 surface expression on human oral squamous carcinoma via PKD2 signal pathway. *Immunobiology*, 217: 385-393. DOI: 10.1016/j.imbio.2011.10.016
- Chen, L., V. Pai, R. Levinson, A.H. Sharpe and G.J. Freeman *et al.*, 2009. Constitutive neuronal expression of the immune regulator, Programmed Death 1 (PD-1), identified during experimental autoimmune uveitis. *Ocul. Immunol. Inflamm.*, 17: 47-55. DOI: 10.1080/09273940802491884
- Cho, H.Y., E.K. Choi, S.W. Lee, K.O. Jung and S.K. Seo *et al.*, 2009. Programmed death-1 receptor negatively regulates LPS-mediated IL-12 production and differentiation of murine macrophage RAW264.7 cells. *Immunol. Lett.*, 127: 39-47. DOI: 10.1016/j.imlet.2009.08.011
- Chou, H.S., C.C. Hsieh, R. Charles, L. Wang and T. Wagner *et al.*, 2012. Myeloid-derived suppressor cells protect islet transplants by B7-H1 mediated enhancement of T regulatory cells. *Transplantation*, 93: 272-282. DOI: 10.1097/TP.0b013e31823ffd39

- Conrad, J.A., R.K. Ramalingam, R.M. Smith, L. Barnett and S.L. Lorey *et al.*, 2011. Dominant clonotypes within HIV-specific T cell responses are programmed death-1<sup>high</sup> and CD127<sup>low</sup> and display reduced variant cross-reactivity. *J. Immunol.*, 186: 6871-6885. DOI: 10.4049/jimmunol.1004234
- Coussens, L.M. and Z. Werb, 2002. Review article inflammation and cancer. *Nature*, 420: 860-867. DOI: 10.1038/nature01322
- D'Souza, M., A.P. Fontenot, D.G. Mack, C. Lozupone and S. Dillon *et al.*, 2007. Programmed death 1 expression on HIV-specific CD4<sup>+</sup> T cells is driven by viral replication and associated with T cell dysfunction. *J. Immunol.*, 179: 1779-1787. PMID: 17641065
- Day, C.L., D.E. Kaufmann, P. Kiepiela, J.A. Brown and E.S. Moodley *et al.*, 2006. PD-1 expression on HIV-specific T cells is associated with T-cell exhaustion and disease progression. *Nature*, 443: 350-354. DOI: 10.1038/nature05115
- Dolganic, A., E. Paek, K. Kodys, J. Thomas and G. Szabo, 2008. Myeloid dendritic cells of patients with chronic HCV infection induce proliferation of regulatory T lymphocytes. *Gastroenterology*, 135: 2119-2127. DOI: 10.1053/j.gastro.2008.07.082
- Dong, H., G. Zhu, K. Tamada and L. Chen, 1999. B7-H1, a third member of the B7 family, co-stimulates T-cell proliferation and interleukin-10 secretion. *Nat. Med.*, 5: 1365-1369. DOI: 10.1038/70932
- Estes, J.D., S.N. Gordon, M. Zeng, A.M. Chahroudi and R.M. Dunham *et al.*, 2008. Early resolution of acute immune activation and induction of PD-1 in SIV-infected sooty mangabeys distinguishes nonpathogenic from pathogenic infection in rhesus macaques. *J. Immunol.*, 180: 6798-6807.
- Finger, L.R., J. Pu, R. Wasserman, R. Vibhakar and E. Louie *et al.*, 1997. The human PD-1 gene: Complete cDNA, genomic organization and developmentally regulated expression in B cell progenitors. *Gene*, 197: 177-187. DOI: 10.1016/S0378-1119(97)00260-6
- Finnefrock, A.C., A. Tang, F. Li, D.C. Freed and M. Feng *et al.*, 2009. PD-1 blockade in rhesus macaques: Impact on chronic infection and prophylactic vaccination. *J. Immunol.*, 182: 980-987.
- Fourcade, J., Z. Sun, M. Benallaoua, P. Guillaume and I.F. Luescher *et al.*, 2010. Upregulation of Tim-3 and PD-1 expression is associated with tumor antigen-specific CD8<sup>+</sup> T cell dysfunction in melanoma patients. *J. Exp. Med.*, 207: 2175-2186. DOI: 10.1084/jem.20100637
- Francisco, L.M., P.T. Sage and A.H. Sharpe, 2010. The PD-1 pathway in tolerance and autoimmunity. *Immunol Rev.*, 236: 219-242. DOI: 10.1111/j.1600-065X.2010.00923.x
- Freel, S.A., K.O. Saunders and G.D. Tomaras, 2011. CD8<sup>+</sup>T-cell-mediated control of HIV-1 and SIV infection. *Immunol. Res.*, 49: 135-146. DOI: 10.1007/s12026-010-8177-7
- Freeman, G.J., E.J. Wherry, R. Ahmed and A.H. Sharpe, 2006. Reinvigorating exhausted HIV-specific T cells via PD-1-PD-1 ligand blockade. *J. Exp. Med.*, 203: 2223-2227. DOI: 10.1084/jem.20061800
- Gary, R., S. Voelkl, R. Palmisano, E. Ullrich and J.J. Bosch *et al.*, 2011. Antigen-specific transfer of functional programmed death ligand 1 from human APCs onto CD8<sup>+</sup> T cells via trogocytosis. *J. Immunol.*, 188: 744-752. DOI: 10.4049/jimmunol.1101412
- Gong, A.Y., R. Zhou, G. Hu, X. Li and P.L. Splinter *et al.*, 2009. MicroRNA-513 regulates B7-H1 translation and is involved in IFN-gamma-induced B7-H1 expression in cholangiocytes. *J. Immunol.*, 182: 1325-1333. PMID: 19155478
- Gotsman, I., A.H. Sharpe and A.H. Lichtman, 2008. T-cell costimulation and coinhibition in atherosclerosis. *Circ. Res.*, 103: 1220-1231. DOI: 10.1161/CIRCRESAHA.108.182428
- Grabie, N., I. Gotsman, R. DaCosta, H. Pang and G. Stavrakis *et al.*, 2007. Endothelial Programmed Death-1 Ligand 1 (PD-L1) regulates CD8<sup>+</sup> T-cell mediated injury in the heart. *Circulation*, 116: 2062-2071. DOI: 10.1161/CIRCULATIONAHA.107.709360
- Ha, S.J., S.N. Mueller, E.J. Wherry, D.L. Barber and R.D. Aubert *et al.*, 2008. Enhancing therapeutic vaccination by blocking PD-1-mediated inhibitory signals during chronic infection. *J. Exp. Med.*, 205: 543-555. DOI: 10.1084/jem.20071949
- He, X.H., L.H. Xu and Y. Liu, 2005. Identification of a novel splice variant of human PD-L1 mRNA encoding an isoform-lacking Igv-like domain. *Acta Pharmacol. Sin.*, 26: 462-468. DOI: 10.1111/j.1745-7254.2005.00086.x
- He, Y.F., G.M. Zhang, X.H. Wang, H. Zhang and Y. Yuan *et al.*, 2004. Blocking programmed death-1 ligand-PD-1 interactions by local gene therapy results in enhancement of antitumor effect of secondary lymphoid tissue chemokine. *J. Immunol.*, 173: 4919-4928. PMID: 15470033

- Hokey, D.A., F.B. Johnson, J. Smith, J.L. Weber and J. Yan *et al.*, 2008. Activation drives PD-1 expression during vaccine-specific proliferation and following lentiviral infection in macaques. *Eur. J. Immunol.*, 38: 1435-1445. DOI: 10.1002/eji.200737857
- Holm, M., F.O. Pettersen and D. Kvale, 2008. PD-1 predicts CD4 loss rate in chronic HIV-1 infection better than HIV RNA and CD38 but not in cryopreserved samples. *Curr. HIV Res.*, 6: 49-58. PMID: 18288975
- Hu, S., D. Tao and P. He, 2004. Expression of costimulatory molecules B7/CD28 in systemic lupus erythematosus. *J. Huazhong Univ. Sci. Technol. Med. Sci.*, 24: 245-265. PMID: 15315338
- Huang, X., F. Venet, Y.L. Wang, A. Lepape and Z. Yuan *et al.*, 2009. PD-1 expression by macrophages plays a pathologic role in altering microbial clearance and the innate inflammatory response to sepsis. *Proc. Natl. Acad. Sci. USA.*, 106: 6303-6308. DOI: 10.1073/pnas.0809422106
- Iglesias-Chiesa, M.C., B. Crabtree-Ramirez and G. Reyes-Teran, 2008. A new therapeutic strategy to control HIV? The PD1 molecule and its role in inhibiting cellular immune responses. *Gac Med. Mex.*, 144: 75-78. PMID: 18619062
- Ishida, M., Y. Iwai, Y. Tanaka, T. Okazaki and G.J. Freeman *et al.*, 2002. Differential expression of PD-L1 and PD-L2, ligands for an inhibitory receptor PD-1, in the cells of lymphohematopoietic tissues. *Immunol. Lett.*, 84: 57-62. PMID: 12161284
- Ishida, Y., Y. Agata, K. Shibahara and T. Honjo, 1992. Induced expression of PD-1, a novel member of the immunoglobulin gene superfamily, upon programmed cell death. *EMBO, J.*, 11: 3887-3895. PMID: 1396582
- Iwamura, K., T. Kato, Y. Miyahara, H. Naota and J. Mineno *et al.*, 2012. siRNA-mediated silencing of PD-1 ligands enhances tumor-specific human T-cell effector functions. *Gene Ther.*, 19: 959-966. DOI: 10.1038/gt.2011.185
- Jin, H.T., A.C. Anderson, W.G. Tan, E.E. West and S.J. Ha *et al.*, 2010. Cooperation of Tim-3 and PD-1 in CD8 T-cell exhaustion during chronic viral infection. *Proc. Natl. Acad. Sci. USA.*, 107: 14733-14738. DOI: 10.1073/pnas.1009731107
- Jin, H.T., Y.H. Jeong, H.J. Park and S.J. Ha, 2011. Mechanism of T cell exhaustion in a chronic environment. *BMB Rep.*, 44: 217-231. DOI: 10.5483/BMBRep.2011.44.4.217
- Jones, R.B., L.C. Ndhlovu, J.D. Barbour, P.M. Sheth and A.R. Jha *et al.*, 2008. Tim-3 expression defines a novel population of dysfunctional T cells with highly elevated frequencies in progressive HIV-1 infection. *J. Exp. Med.*, 205: 2763-2779. DOI: 10.1084/jem.20081398
- Karwacz, K., F. Arce, C. Bricogne, G. Kochan and D. Escors, 2012. PD-L1 co-stimulation, ligand-induced TCR down-modulation and anti-tumor immunotherapy. *Oncoimmunology*, 1: 86-88. DOI: 10.4161/onci.1.1.17824
- Kassu, A., R.A. Marcus, M.B. D'Souza, E.A. Kelly-McKnight and L. Golden-Mason *et al.*, 2010. Regulation of virus-specific CD4<sup>+</sup> T cell function by multiple costimulatory receptors during chronic HIV infection. *J. Immunol.*, 185: 3007-3018. DOI: 10.4049/jimmunol.1000156
- Keir, M.E., Y.E. Latchman, G.J. Freeman and A.H. Sharpe, 2005. Programmed Death-1 (PD-1):PD-ligand 1 interactions inhibit TCR-mediated positive selection of thymocytes. *J. Immunol.*, 175: 7372-7279. PMID: 16301644
- Kinter, A.L., E.J. Godbout, J.P. McNally, I. Sereti and G.A. Roby *et al.*, 2008. The common gamma-chain cytokines IL-2, IL-7, IL-15 and IL-21 induce the expression of programmed death-1 and its ligands. *J. Immunol.*, 181: 6738-6746. PMID: 18981091
- Latchman, Y., C.R. Wood, T. Chernova, D. Chaudhary and M. Borde *et al.*, 2001. PD-L2 is a second ligand for PD-1 and inhibits T cell activation. *Nat. Immunol.*, 2: 261-268. DOI: 10.1038/85330
- Lazar-Molnar, E., Q. Yan, E. Cao, U. Ramagopal and S.G. Nathanson *et al.*, 2008. Crystal structure of the complex between Programmed Death-1 (PD-1) and its ligand PD-L2. *Proc. Natl. Acad. Sci. USA.*, 105: 10483-10488. DOI: 10.1073/pnas.0804453105
- Lee, H., J.H. Kim, S.Y. Yang, J. Kong and M. Oh *et al.*, 2010. Peripheral blood gene expression of B7 and CD28 family members associated with tumor progression and microscopic lymphovascular invasion in colon cancer patients. *J. Cancer Res. Clin. Oncol.*, 136: 1445-1452. DOI: 10.1007/s00432-010-0800-4
- Liang, M., H. Yang and J. Fu, 2009. Nimesulide inhibits IFN- $\gamma$ -induced programmed death-1-ligand 1 surface expression in breast cancer cells by COX-2 and PGE<sub>2</sub> independent mechanisms. *Cancer Lett.*, 276: 47-52. DOI: 10.1016/j.canlet.2008.10.028
- Lipp, M., C. Brandt, F. Dehghani, E. Kwidzinski and I. Bechmann, 2007. PD-L1 (B7-H1) regulation in zones of axonal degeneration. *Neurosci. Lett.*, 425: 156-161. DOI: 10.1016/j.neulet.2007.07.053

- Mages, H.W., A. Hutloff, C. Heuck, K. Buchner and H. Himmelbauer *et al.*, 2000. Molecular cloning and characterization of murine ICOS and identification of B7h as ICOS ligand. *Eur. J. Immunol.*, 30: 1040-1047. PMID: 10760791
- Manches, O., D. Munn, A. Fallahi, J. Lifson and L. Chaperot *et al.*, 2008. HIV-activated human plasmacytoid DCs induce Tregs through an indoleamine 2,3-dioxygenase-dependent mechanism. *J. Clin. Invest.*, 118: 3431-3439. DOI: 10.1172/JCI34823
- Mazanet, M.M. and C.C. Hughes, 2002. B7-H1 is expressed by human endothelial cells and suppresses T cell cytokine synthesis. *J. Immunol.*, 169: 3581-3588. PMID: 12244148
- Meier, A., A. Bagchi, H.K. Sidhu, G. Alter and T.J. Suscovich *et al.*, 2008. Upregulation of PD-L1 on monocytes and dendritic cells by HIV-1 derived TLR ligands. *AIDS*, 22: 655-658. DOI: 10.1097/QAD.0b013e3282f4de23
- Menke, J., J.A. Lucas, G.C. Zeller, M.E. Keir and X.R. Huang *et al.*, 2007. Programmed Death 1 Ligand (PD-L) 1 and PD-L2 limit autoimmune kidney disease: Distinct roles. *J. Immunol.*, 179: 7466-7477. PMID: 18025191
- Messal, N., N.E. Serriari, S. Pastor, J.A. Nunes and D. Olive, 2011. PD-L2 is expressed on activated human T cells and regulates their function. *Mol. Immunol.*, 48: 2214-2219. DOI: 10.1016/j.molimm.2011.06.436
- Moll, M., C. Kuylenstierna, V.D. Gonzalez, S.K. Andersson and L. Bosnjak *et al.*, 2009. Severe functional impairment and elevated PD-1 expression in CD1d-restricted NKT cells retained during chronic HIV-1 infection. *Eur. J. Immunol.*, 39: 902-911. DOI: 10.1002/eji.200838780
- Muthumani, K., A.Y. Choo, D.J. Shedlock, D.J. Laddy and S.G. Sundaram *et al.*, 2008. Human immunodeficiency virus type 1 Nef induces programmed death 1 expression through a p<sup>38</sup> mitogen-activated protein kinase-dependent mechanism. *J. Virol.*, 82: 11536-11544. DOI: 10.1128/JVI.00485-08
- Muthumani, K., D.J. Shedlock, D.K. Choo, P. Fagone and O.U. Kawalekar *et al.*, 2011. HIV-mediated phosphatidylinositol 3-kinase/serine-threonine kinase activation in APCs leads to programmed death-1 ligand upregulation and suppression of HIV-specific CD8 T cells. *J. Immunol.*, 187: 2932-2943. DOI: 10.4049/jimmunol.1100594
- Nakae, S., H. Suto, M. Iikura, M. Kakurai and J.D. Sedgwick *et al.*, 2006. Mast cells enhance T cell activation: Importance of mast cell costimulatory molecules and secreted TNF. *J. Immunol.*, 176: 2238-2248. PMID: 16455980
- Nakanjako, D., I. Ssewanyana, H. Mayanja-Kizza, A. Kiragga and R. Colebunders *et al.*, 2011. High T-cell immune activation and immune exhaustion among individuals with suboptimal CD4 recovery after 4 years of antiretroviral therapy in an African cohort. *BMC Infect. Dis.*, 11: 43-43. DOI: 10.1186/1471-2334-11-43
- Ni, R., K. Ihara, K. Miyako, R. Kuromaru and M. Inuo *et al.*, 2007. PD-1 gene haplotype is associated with the development of type 1 diabetes mellitus in Japanese children. *Hum. Genet.*, 121: 223-232. DOI: 10.1007/s00439-006-0309-8
- Nurieva, R., S. Thomas, T. Nguyen, N. Martin-Orozco and Y. Wang *et al.*, 2006. T-cell tolerance or function is determined by combinatorial costimulatory signals. *EMBO J.*, 25: 2623-2633. DOI: 10.1038/sj.emboj.7601146
- Okazaki, T., A. Maeda, H. Nishimura, T. Kurosaki and T. Honjo, 2001. PD-1 immunoreceptor inhibits B cell receptor-mediated signaling by recruiting src homology 2-domain-containing tyrosine phosphatase 2 to phosphotyrosine. *Proc. Natl. Acad. Sci. USA.*, 98: 13866-13871. DOI: 10.1073/pnas.231486598
- Parry, R.V., J.M. Chemnitz, K.A. Frauwirth, A.R. Lanfranco and I. Braunstein *et al.*, 2005. CTLA-4 and PD-1 receptors inhibit T-cell activation by distinct mechanisms. *Mol. Cell Biol.*, 25: 9543-9553. DOI: 10.1128/MCB.25.21.9543-9553.2005
- Paterson, A.M., V.K. Vanguri and A.H. Sharpe, 2009. SnapShot: B7/CD28 costimulation. *Cell*, 137: 974-974. DOI: 10.1016/j.cell.2009.05.015
- Pentcheva-Hoang, T., L. Chen, D.M. Pardoll and J.P. Allison, 2007. Programmed death-1 concentration at the immunological synapse is determined by ligand affinity and availability. *Proc. Natl. Acad. Sci. USA.*, 104: 17765-17770. DOI: 10.1073/pnas.0708767104
- Petroff, M.G. and A. Perchellet, 2010. B7 family molecules as regulators of the maternal immune system in pregnancy. *Am. J. Reprod. Immunol.*, 63: 506-519. DOI: 10.1111/j.1600-0897.2010.00841.x
- Petrovas, C., J.P. Casazza, J.M. Brenchley, D.A. Price and E. Gostick *et al.*, 2006. PD-1 is a regulator of virus-specific CD8<sup>+</sup> T cell survival in HIV infection. *J. Exp. Med.*, 203: 2281-2292. DOI: 10.1084/jem.20061496

- Porichis, F. and D.E. Kaufmann, 2011. HIV-specific CD4 T cells and immune control of viral replication. *Curr. Opin. HIV AIDS*, 6: 174-180. DOI: 10.1097/COH.0b013e3283454058
- Porichis, F. and D.E. Kaufmann, 2012. Role of PD-1 in HIV pathogenesis and as target for therapy. *Curr. HIV/AIDS Rep.*, 9: 81-90. DOI: 10.1007/s11904-011-0106-4
- Porichis, F., D.S. Kwon, J. Zupkosky, D.P. Tighe and A. McMullen *et al.*, 2011. Responsiveness of HIV-specific CD4 T cells to PD-1 blockade. *Blood*, 118: 965-674. DOI: 10.1182/blood-2010-12-328070
- Quigley, M., F. Pereyra, B. Nilsson, F. Porichis and C. Fonseca *et al.*, 2010. Transcriptional analysis of HIV-specific CD8<sup>+</sup> T cells shows that PD-1 inhibits T cell function by upregulating BATF. *Nat. Med.*, 16: 1147-1151. DOI: 10.1038/nm.2232
- Radziewicz, H., C.C. Ibegbu, M.L. Fernandez, K.A. Workowski and K. Obideen *et al.*, 2007. Liver-infiltrating lymphocytes in chronic human hepatitis C virus infection display an exhausted phenotype with high levels of PD-1 and low levels of CD127 expression. *J. Virol.*, 81: 2545-2553. DOI: 10.1128/JVI.02021-06
- Rajasalu, T., H. Brosi, C. Schuster, A. Spyranitis and B.O. Boehm *et al.*, 2010. Deficiency in B7-H1 (PD-L1)/PD-1 coinhibition triggers pancreatic  $\beta$ -cell destruction by insulinspecific, murine CD8 T-cells. *Diabetes*, 59: 1966-1973. DOI: 10.2337/db09-1135
- Raptopoulou, A.P., G. Bertias, D. Makrygiannakis, P. Verginis and I. Kritikos *et al.*, 2010. The programmed death 1/programmed death ligand 1 inhibitory pathway is up-regulated in rheumatoid synovium and regulates peripheral T cell responses in human and murine arthritis. *Arthritis Rheum*, 62: 1870-1880. DOI: 10.1002/art.27500
- Riley, J.L., 2009. PD-1 signaling in primary T cells. *Immunol. Rev.*, 229: 114-125. DOI: 10.1111/j.1600-065X.2009.00767.x
- Rodrigue-Gervais, I.G., H. Rigsby, L. Jouan, D. Saue and R.P. Sekaly *et al.*, 2010. Dendritic cell inhibition is connected to exhaustion of CD8<sup>+</sup> T cell polyfunctionality during chronic hepatitis C virus infection. *J. Immunol.*, 184: 3134-3144. DOI: 10.4049/jimmunol.0902522
- Rodriguez-Garcia, M., F. Porichis, O.G.D. Jong, K. Levi and T.J. Diefenbach *et al.*, 2011. Expression of PD-L1 and PD-L2 on human macrophages is up-regulated by HIV-1 and differentially modulated by IL-10. *J. Leukoc Biol.*, 89: 507-515. DOI: 10.1189/jlb.0610327
- Rosignoli, G., A. Cranage, C. Burton, M. Nelson and A. Steel *et al.*, 2007. Expression of PD-L1, a marker of disease status, is not reduced by HAART in aviraemic patients. *AIDS*, 21: 1379-1381. DOI: 10.1097/QAD.0b013e3281de7296
- Rosignoli, G., C.H. Lim, M. Bower, F. Gotch and N. Imami, 2009. Programmed Death (PD)-1 molecule and its ligand PD-L1 distribution among memory CD4 and CD8 T cell subsets in human immunodeficiency virus-1-infected individuals. *Clin. Exp. Immunol.*, 157: 90-97. DOI: 10.1111/j.1365-2249.2009.03960.x
- Said, E.A., F.P. Dupuy, L. Trautmann, Y. Zhang and Y. Shi *et al.*, 2010. Programmed death-1-induced interleukin-10 production by monocytes impairs CD4<sup>+</sup> T cell activation during HIV infection. *Nat. Med.*, 16: 452-459. DOI: 10.1038/nm.2106
- Sauce, D., J.R. Almeida, M. Larsen, L. Haro and B. Autran *et al.*, 2007. PD-1 expression on human CD8 T cells depends on both state of differentiation and activation status. *AIDS*, 21: 2005-2013. DOI: 10.1097/QAD.0b013e3282eee548
- Schwartz, J.C.D., X. Zhang, A.A. Fedorov, S.G. Nathenson and S.C. Almo, 2001. Structural basis for co-stimulation by the human CTLA-4/B7-2 complex. *Nature*, 410: 604-608. DOI: 10.1038/35069112
- Sheppard, K.A., L.J. Fitts, J.M. Lee, C. Benander and J.A. George *et al.*, 2004. PD-1 inhibits T-cell receptor induced phosphorylation of the ZAP70/CD3 $\zeta$  signalosome and downstream signaling to PKC $\theta$ . *FEBS Lett.*, 574: 37-41. DOI: 10.1016/j.febslet.2004.07.083
- Shetty, R.D., V. Velu, K. Titanji, S.E. Bosinger and G.J. Freeman *et al.*, 2012. PD-1 blockade during chronic SIV infection reduces hyperimmune activation and microbial translocation in rhesus macaques. *J. Clin. Invest.*, 122: 1712-1716. DOI: 10.1172/JCI60612
- Shin, H., S.D. Blackburn, A.M. Intlekofer, C. Kao and J.M. Angelosanto *et al.*, 2009. A role for the transcriptional repressor Blimp-1 in CD8<sup>+</sup> T cell exhaustion during chronic viral infection. *Immunity*, 31: 309-320. DOI: 10.1016/j.immuni.2009.06.019
- Shinohara, T., M. Taniwaki, Y. Ishida, M. Kawaichi and T. Honjo, 1994. Structure and chromosomal localization of the human PD-1 gene (PDCD1). *Genomics*, 23: 704-706. DOI: 10.1006/geno.1994.1562

- Stanciu, L.A., C.M. Bellettato, V. Laza-Stanca, A.J. Coyle and A. Papi *et al.*, 2006. Expression of Programmed Death-1 Ligand (PD-L) 1, PD-L2, B7-H3 and inducible costimulator ligand on human respiratory tract epithelial cells and regulation by respiratory syncytial virus and type 1 and 2 cytokines. *J. Infect. Dis.*, 193: 404-412. DOI: 10.1086/499275
- Swain, S.L., K.K. McKinstry and T.M. Strutt, 2012. Expanding roles for CD4<sup>+</sup> T cells in immunity to viruses. *Nat. Rev. Immunol.*, 12: 136-148. DOI: 10.1038/nri3152
- Tendeiro, R., R.B. Foxall, A.P. Baptista, F. Pinto and R.S. Soares *et al.*, 2012. PD-1 and its ligand PD-L1 are progressively up-regulated on CD4 and CD8 T-cells in HIV-2 infection irrespective of the presence of viremia. *AIDS*, 26: 1065-1071. DOI: 10.1097/QAD.0b013e32835374db
- Trautmann, L., L. Janbazian, N. Chomont, E.A. Said and S. Gimmig *et al.*, 2006. Upregulation of PD-1 expression on HIV-specific CD8<sup>+</sup> T cells leads to reversible immune dysfunction. *Nat. Med.*, 12: 1198-1202. DOI: 10.1038/nm1482
- Tseng, S.Y., M. Otsuji, K. Gorski, X. Huang and J.E. Slansky *et al.*, 2001. B7-DC, a new dendritic cell molecule with potent costimulatory properties for T cells. *J. Exp. Med.*, 193: 839-846. PMID: 11283156
- Urbani, S., B. Amadei, D. Tola, M. Massari and S. Schivazappa *et al.*, 2006. PD-1 expression in acute hepatitis C virus (HCV) infection is associated with HCV-specific CD8 exhaustion. *J. Virol.*, 80: 11398-11403. DOI: 10.1128/JVI.01177-06
- Velu, V., K. Titanji, B. Zhu, S. Husain and A. Pladevega *et al.*, 2009. Enhancing SIV-specific immunity *in vivo* by PD-1 blockade. *Nature*, 458: 206-210. DOI: 10.1038/nature07662
- Venkatachari, N.J., W.G. Buchanan and V. Ayyavoo, 2008. Human Immunodeficiency Virus (HIV-1) infection selectively downregulates PD-1 expression in infected cells and protects the cells from early apoptosis *in vitro* and *in vivo*. *Virology*, 376: 140-153. DOI: 10.1016/j.virol.2008.03.015
- Vibhakar, R., G. Juan, F. Traganos, Z. Darzynkiewicz and L.R. Finger, 1997. Activation-induced expression of human programmed death-1 gene in T-lymphocytes. *Exp. Cell Res.*, 232: 25-28. DOI: 10.1006/excr.1997.3493
- Virgin, H.W., E.J. Wherry and R. Ahmed, 2009. Redefining chronic viral infection. *Cell*, 138: 30-50. DOI: 10.1016/j.cell.2009.06.036
- Wagner, C.J., S. Huber, S. Wirth and D. Voehringer, 2010. Chitin induces upregulation of B7-H1 on macrophages and inhibits T-cell proliferation. *Eur. J. Immunol.*, 40: 2882-2890. DOI: 10.1002/eji.201040422
- Wells, A.D., 2009. New insights into the molecular basis of T cell anergy: Anergy factors, avoidance sensors and epigenetic imprinting. *J. Immunol.*, 182: 7331-7341. DOI: 10.4049/jimmunol.0803917
- Wherry, E.J. and R. Ahmed, 2004. Memory CD8 T-cell differentiation during viral infection. *J. Virol.*, 78: 5535-5545. DOI: 10.1128/JVI.78.11.5535-5545.2004
- Wiendl, H., O. Neuhaus, M. Mehling, S. Wintterle and B. Schreiner *et al.*, 2003. The CD28 related molecule ICOS: T cell modulation in the presence and absence of B7.1/2 and regulational expression in multiple sclerosis. *J. Neuroimmunol.*, 140: 177-187. DOI: 10.1016/S0165-5728(03)00172-3
- Wolfe, S.J., J. Strebovsky, H. Bartz, A. Sahr and C. Arnold *et al.*, 2011. PD-L1 expression on tolerogenic APCs is controlled by STAT-3. *Eur. J. Immunol.*, 41: 413-424. DOI: 10.1002/eji.201040979
- Wong, R.M., K.A. Smith, V.L. Tam, R.R. Pagarigan and B.L. Meisenburg *et al.*, 2009. TLR-9 signaling and TCR stimulation co-regulate CD8<sup>+</sup> T cell-associated PD-1 expression. *Immunol. Lett.*, 127: 60-67. DOI: 10.1016/j.imlet.2009.09.002
- Yamamoto, T., D.A. Price, J.P. Casazza, G. Ferrari and M. Nason *et al.*, 2011. Surface expression patterns of negative regulatory molecules identify determinants of virus-specific CD8<sup>+</sup> T-cell exhaustion in HIV infection. *Blood*, 117: 4805-4815. DOI: 10.1182/blood-2010-11-317297
- Youngblood, B., E.J. Wherry and R. Ahmed, 2012. Acquired transcriptional programming in functional and exhausted virus-specific CD8 T cells. *Curr. Opin. HIV AIDS*, 7: 50-57. DOI: 10.1097/COH.0b013e32834ddef2
- Youngblood, B., K.J. Oestreich, S.J. Ha, J. Duraiswamy and R.S. Akondy *et al.*, 2011. Chronic virus infection enforces demethylation of the locus that encodes PD-1 in antigen-specific CD8<sup>+</sup> T cells. *Immunity*, 35: 400-412. DOI: 10.1016/j.immuni.2011.06.015
- Youngnak, P., Y. Kozono, H. Kozono, H. Iwai and N. Otsuki *et al.*, 2003. Differential binding properties of B7-H1 and B7-DC to programmed death-1. *Biochem. Biophys. Res. Commun.*, 307: 672-677. DOI: 10.1016/S0006-291X(03)01257-9

- Youngnak-Piboonratanakit, P., F. Tsushima, N. Otsuki, H. Igarashi and U. Machida *et al.*, 2004. The expression of B7-H1 on keratinocytes in chronic inflammatory mucocutaneous disease and its regulatory role. *Immunol. Lett.*, 94: 215-222. DOI: 10.1016/j.imlet.2004.05.007
- Zajac, A.J., J.N. Blattman, K. Murali-Krishna, D.J. Sourdive and M. Suresh *et al.*, 1998. Viral immune evasion due to persistence of activated T cells without effector function. *J. Exp. Med.*, 188: 2205-2213. PMID: 9858507
- Zhang, J.Y., Z. Zhang, X. Wang, J.L. Fu and J. Yao *et al.*, 2007. PD-1 up-regulation is correlated with HIV-specific memory CD8<sup>+</sup> T-cell exhaustion in typical progressors but not in long-term nonprogressors. *Blood*, 109: 4671-4678. DOI: 10.1182/blood-2006-09-044826
- Zhang, M., X.Q. Xiao, Y.F. Jiang, Y.S. Liang and M.Y. Peng *et al.*, 2011. DNA demethylation in PD-1 gene promoter induced by 5-azacytidine activates PD-1 expression on Molt-4 cells. *Cell Immunol.*, 271: 450-454. DOI: 10.1016/j.cellimm.2011.08.014
- Zhang, X., J.C. Schwartz, X. Guo, S. Bhatia and E. Cao *et al.*, 2004. Structural and functional analysis of the costimulatory receptor programmed death-1. *Immunity*, 20: 337-347. DOI: 10.1016/S1074-7613(04)00051-2