

Induced Production of α -Tocotrienol by Co-Suppression of Tocopherol Cyclase Gene in Vegetable Crops

^{1,2}Umaiya Munusamy, ^{1,3}Siti Nor Akmar Abdullah,
^{1,3}Maheran Abdul Aziz, ⁴Huzwah Khazaai and ¹Lai Mun Seong

¹Department of Agriculture Technology, Faculty of Agriculture,
University Putra Malaysia, 43400 UPM Serdang, Selangor Darul Ehsan, Malaysia

²Institute of Biological Sciences, Faculty of Science, University of Malaya, 50603 Kuala Lumpur, Malaysia

³Laboratory of Plantation Crops, Institute of Tropical Agriculture,

⁴Department of Biomedical Science, Faculty of Medicine and Health Sciences,
University Putra Malaysia, 43400 UPM Serdang, Selangor Darul Ehsan, Malaysia

Received 2013-09-01; Revised 2013-10-01; Accepted 2013-10-16

ABSTRACT

This study involves the induced production of α -tocotrienol by the co-suppression of tocopherol cyclase gene. The Tocopherol/Tocotrienol Cyclase (TC) is a key enzyme involved in the biosynthesis of α -tocotrienol. The partial conserved cDNA of TC gene isolated from the oil palm (*Elaeis guineensis*) that shared greater than 75% sequence identity with TC sequences from other plant species was used to silence TC gene expression in *Amaranthus* sp. and *Allium porrum* leaves by co-suppression strategies. This was carried out using recombinant vector constructs designated as p4a11 and p4c9 containing TC driven by the maize ubiquitin promoter (Ubi1P) and the oil palm leaf-specific promoter (LHCB), respectively. In the *Amaranthus* sp. leaves infiltrated with p4a11, the α -tocopherol suppression was 85.0, 71.0 and 74.0% on day three, five and seven, respectively and complete suppression was observed in the leaves infiltrated with p4c9. Whereas, only 11.0, 7.0 and 29.0% suppression was detected in *Allium porrum* leaves infiltrated with p4a11 and 45.0, 4.0 and 57.0% of suppression was observed in *Allium porrum* infiltrated with p4c9 on day three, five and seven, respectively. Silencing of TC gene driven by LHCB (p4c9) causes the reduction of α -tocopherol production better than by Ubi1P (p4a11). The results have demonstrated that gene silencing through co-suppression is capable of producing α -tocotrienol. However, there is no correlation between the α -tocotrienol production and the α -tocopherol suppression. It has been suggested that the transgene mediated co-suppression mechanism had triggered the vitamin E biosynthesis metabolic pathway to shift towards producing α -tocotrienol.

Keywords: Vitamin E Biosynthetic Pathway, Co-Suppression, Tocopherol Cyclase, α -Tocopherol, α -Tocotrienol, Tissue-Specific Promoter

1. INTRODUCTION

Vitamin E refers to four isomers (α , β , γ and δ) each of tocopherol and tocotrienol produced by the shikimate and non-mevalonate pathways (Fig. 1). Both the α -tocopherol and α -tocotrienol have high antioxidant

properties and play an important role in influencing the biological activities in human beings (Traber and Atkinson, 2007). In addition, it has been reported that the α -tocotrienol has several beneficial effects on human health better than the α -tocopherol due to the shorter and thicker prenyl tail that makes the molecule more flexible

Corresponding Author: Siti Nor Akmar Abdullah, Department of Agriculture Technology, Faculty of Agriculture,
University Putra Malaysia, 43400 UPM Serdang, Selangor Darul Ehsan, Malaysia

to penetrate the cell membranes. This results in superior biological activities that help to promote cellular growth (Nesaretnam *et al.*, 1995) and support the body's innate neuroprotective characteristics (Sanagi *et al.*, 2006; Ryyanen *et al.*, 2004).

Plants widely differ in their vitamin E isomers level and no single plant provides the entire vitamin E family of isomers in nutritionally adequate amounts. Even within a plant, large variances exist in the proportions of the α , β , γ and δ isomers. Typically, green vegetables have high level of tocopherol and certain plant seeds contain high level of tocotrienol (Ajjawi and Shintani, 2004; Abbasi *et al.*, 2007). Therefore, plants that have been metabolically engineered to produce both compounds in vegetative tissues are attractive sources for both the tocopherol and tocotrienol.

Tocopherol Cyclase (TC) adds a second ring at the junction between the aromatic head group and the prenyl

tail to create a two-ring structure which is known as chromanol ring in the tocopherol biosynthesis (Ajjawi and Shintani, 2004). Subsequent methylation of MPBQ methyltransferase and γ -tocopherol methyltransferase produce β and α -tocochromanols. Metabolite engineering to modify vitamin E content and composition had been carried out in potato with TC gene (Hofius *et al.*, 2004). In addition, genetic engineering and pathway modification through suppression was applied in *petunia* with CHS gene (Napoli *et al.*, 1990), tobacco with *AtFAD7* gene (Oka *et al.*, 2010) and in tomato with *GA 20-oxidase1* gene (Olimpieri *et al.*, 2011). The above investigations have concluded that when one of the metabolite present in the biosynthesis pathway is suppressed, the other pathway that exists will be initiated to synthesize other forms of metabolites that are necessary for the survival of the plants.

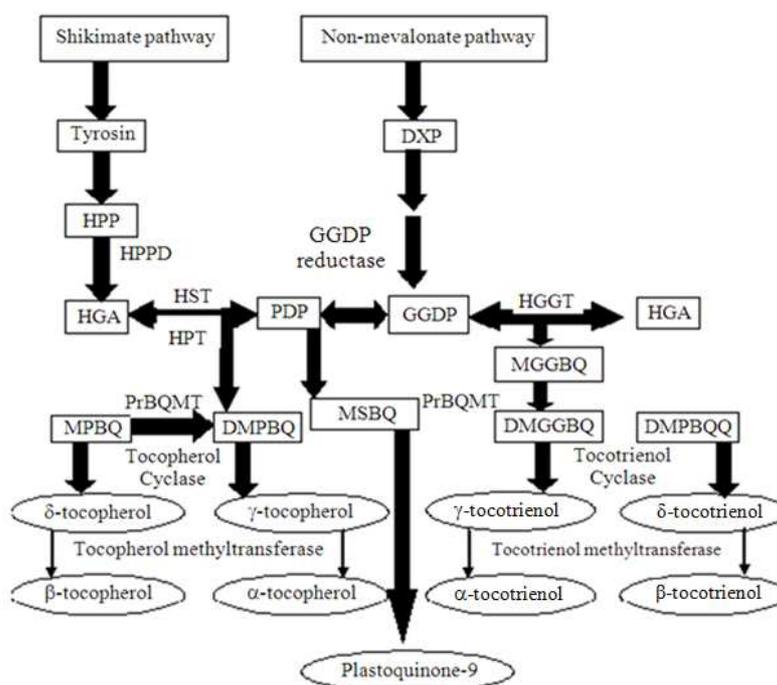


Fig. 1. The tocopherols and plastoquinone biosynthesis pathway in plants. It represents the enzymatic reactions and the intermediate products that are involved. Shikimate and non-mevalonate pathway takes place in the plastid and the formed metabolites are transported to other part of the plant. Homogentisate Acid (HGA) and Phytidyldiphosphate (PDP), derived from cytosolic aromatic amino acid metabolism and plastidic Deoxyxylulose 5-Phosphate Pathway (DXP), respectively serve as substrates for the synthesis of tocopherol. Reactions between HGA and PDP or HGA with GGDP carry out by Homogentisate Phytyltransferase (HPT) or Homogentisate Geranylgeranyltransferase (HGGT) will produce tocopherol. The intermediate products 2-Methyl-6-Phytinyl-Benzoquinol (DMPBQ) and 2-Methyl-6-Geranylgeranyl-Benzoquinol (MGGBQ) will be further cyclized and methylated to produce α -tocopherol. Homogentisate Solanesyltransferase (HST) catalyzes the prenylation and decarboxylation of HGA to form 2-Methyl-6-Solanesyl-1,4-Benzoquinol (MSBQ). Enzymes in BOLD and UPPER CASE are specific to the synthesis of the subsequent intermediate products

Partial conserved cDNA that encodes the open reading frame of TC gene was isolated and was expressed with Ubiquitin Promoter from maize (Ubi1P) or leaf-specific promoter from oil palm (LHCB) using the pDRB6b expression vector. Since most of the agricultural crops are not amenable to stable genetic transformation, agro-infiltration based system was used as a tool for transferring the transformed *Agrobacterium* with the foreign gene directly into the intact leaf tissues of *Amaranthus* sp. and *Allium porrum*. Suppression level of α -tocopherol as well as the formation of α -tocotrienol was successfully studied using these two different plant promoters. The aim of the present study is to induce the production of α -tocotrienol in leaves tissue that do not contain of α -tocotrienol by the co-suppression of TC.

2. MATERIALS AND METHODS

2.1. Gene Constructs

Recombinant vector constructs for transient gene expression and the control vectors were prepared according to the standard procedures (Sambrook *et al.*, 1989). The PCR and DNA sequencing primers used in this study are shown in **Table 1** and the bacterial strains and the plasmids are listed in **Table 2**. The competent cells of *E. coli* DH5 α were purchased from Yeastern Biotech Co., Ltd, Shijr Taipei, Taiwan. The pDRB6b vector digested with *HindIII/KpnI*, served as the backbone vector for p4a11 and p4c9. Ubiquitin promoter was isolated by using *HindIII/BglII* restriction enzyme from the pDRB8 vector while the LHCB was isolated by amplifying the sequences using FLHCB and RLHCB primers from the pGWLSO1 vector with the similar restriction site. The pDRB8 vector was also digested to isolate the NosT using *HindIII/KpnI*. Tocopherol Cyclase gene (TC) was amplified with FtcB2/RtcH2 primers from γ T&A-TC plasmid to produce *BglII/HindIII* ends. The following amplification program was used: Heating at 95°C for 5 min for initial denaturation; 35 cycles using a Biometra®Tpersonal Thermocycler (Goettingen, Germany) at 94°C for 30 s for a second denaturation process, annealing temperature was optimised at 4°C lesser than its melting temperature; 72°C at 45 s for DNA extension; 72°C at 5 min for final DNA extension and stored at 4°C. The TC gene was ligated to Ubi1P having *BglII* restriction sites at 3' end to NosT having *HindIII* restriction sites at its 5' end. The resultant plasmid was named p4a11 (**Fig. 2a**). For p4c9 (**Fig.**

2b), the LHCB was ligated to TC gene and with NosT fragment having similar restriction sites as in p4a11.

The isolated recombinant vector constructs were linearised using single enzyme digestion according to the manufacturer's recommendations. The sizes of the produce vector constructs were compared with the control vectors. The successfully produced constructs were further confirmed by amplifying the TC gene with primary and nested primers (vcFTC, vcRTC, npfTC) that were flanked by Ubi1P or LHCB and NosT fragment between LB and RB2 border. The amplified PCR product was sequenced and analysed using the NCBI Blast.

2.2. Selection and Growth of Transformed Bacteria

E. coli DH5 α was grown for 16 h at 37°C (Sadre *et al.*, 2006). *Agrobacterium* strains (AGL1) was obtained from Dr N.M. Upadhyaya, CSIRO, Australia. The AGL1 carrying the plasmid (pDRB6b, pDRB5b, p4a11 and p4c9) was streaked on LB agar (Merck, Darmstadt, Germany) plates containing spectinomycin (Calbiochem® Germany) and rifampicin (Calbiochem® Canada). The plates were incubated at 28°C for two days. The AGL1 was inoculated in 5 mL LB broth with the same antibiotics and was grown overnight at 28°C on a shaker at 150 rpm (Lee and Yang, 2006). All the transformed bacterial strains were grown on Luria-Bertani (LB) agar or broth (Difco™, USA). Broth culture(s) were grown with shaking at 200 rpm.

2.3. Agro-Infiltration of Vegetable Leaves

This section was performed as described by Lee and Yang (2006) with only slight modification. Two milliliter of culture was collected in a sterile microcentrifuge by centrifugation at 8000 rpm for 10 min. The *Agrobacteria* pellet was resuspended in 1 mL sterilised induction medium (10.5 g L⁻¹ K₂HPO₄, 4.5 g L⁻¹ KH₂PO₄, 1 g L⁻¹ (NH₄)₂SO₄, 0.5 g L⁻¹ NaCitrate, 1 g L⁻¹ glucose, 1 g L⁻¹ fructose, 4 g L⁻¹ glycerol, 1 mM MgSO₄, 10 mM MES, pH 5.6).

Four milliliter of sterilised infiltration medium (10 mM MgSO₄, 10 mM MES, pH 5.6) was added to make up 5 mL of medium supplemented with 100 μ M acetosyringone (Sigma Aldrich® Chemistry, USA) and 50 mg L⁻¹ of spectinomycin. The *Agrobacteria* suspension was further grown at 28°C for 6 h. The bacterial cultures were collected by centrifugation and were resuspended in infiltration medium containing 200 μ M acetosyringone to 1.0 OD at 600 nm.

Table 1. PCR and DNA sequencing primers used in this study

Primer name	T _m °C	Nucleotide sequence 5' to 3'	Template
FtcB2	61.7	TTAATA <i>AGATCT</i> AGGTCGAAGTATGTCC	yT&A-TC
RtcH2	59.0	AATAAAATTA <i>AGCTTTT</i> CCCACAATTGC	yT&A-TC
vcFTC	66.5	ACAGCAGGCTGGCTGCAGC	p4a11, p4c9
vcRTC	66.5	TTGTTGGAGCCC GCAAGGGG	p4a11, p4c9
npfTC	64.5	TGCATGGCAGCTGGCTTGTC	p4a11, p4c9
FLHCB	63.1	AGGGGGA <i>AGCTT</i> AAATAAAAAGATCGG	pGWLSO1
RLHCB	63.1	TTCAAT <i>AGATCT</i> GATGCCTTGGAG	pGWLSO1

*Sequence in italics underlined represents restriction enzyme sites

Table 2. Bacterial strains and plasmids used in this study.

	Genotype/relevant properties	Source/reference
<i>Strains</i>		
<i>Escherichia coli</i> DH5a	<i>endA1 recA1 relA1 gyrA96 hsdR17(r_k⁻, m_k⁺) phoA supE44 thi-1 Δ(lacZYA-argF)U169Φ80Δ(lacZ)M15F-</i>	Yeastern Biotech Co.,
<i>Agrobacterium tumefaciens</i> (AGL1)	Broad host range strain for plant transformation, Rif ^R	CSIRO Plant Industry
<i>Plasmids</i>		
pDRB5b	a donor of Ubi1-intron, <i>uidA</i> , linearised with <i>KpnI</i>	CSIRO Plant Industry
pDRB6b	An expression vector, Spec ^R , linearised with <i>KpnI</i>	CSIRO Plant Industry
pDRB8	a donor of Ubi1P and NosT, linearised with <i>SmaI</i>	CSIRO Plant Industry
pGWLSO1	a donor of leaf specific promoter (LHCB)	MPOB
yT&A	cloning vector, Amp ^R	Yeastern Biotech Co.,
yT&A-TC	recombinant vector, a donor of TC gene	This study
p4a11	recombinant vector, T-DNA region consist of Ubi1P, TC, NosT, Spec ^R , linearised with <i>NotI</i>	This study
p4c9	recombinant vector, T-DNA region consist of LHCB, TC, NosT, Spec ^R , linearised with <i>NotI</i>	This study

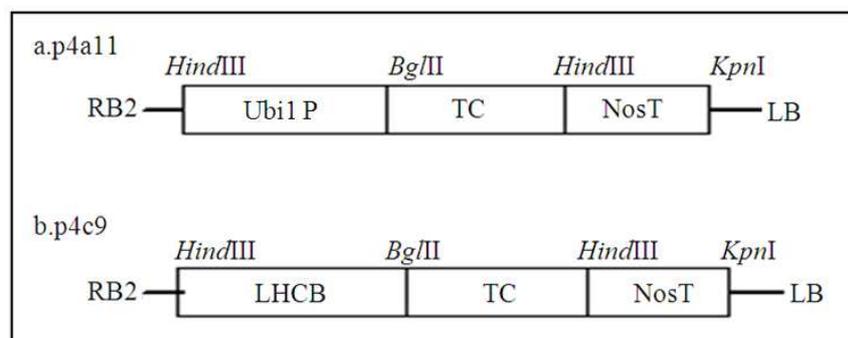


Fig. 2. Chimeric recombinant vector constructs with Tocopherol Cyclase gene (TC). RB2: Second right border, LB: Left Border. Gene constructs for transient gene expression were prepared by fusing together partial sense sequences for TC (622 bp) with maize ubiquitin promoter-Ubi1P (950 bp) or leaf-specific promoter-LHCB (1000 bp), NosT indicates the nos terminator (503 bp). The restriction enzyme sites in italics were used for ligations. The length of (a) p4a11~2075 bp and (b) p4c9~2125 bp

Agroinfiltration was conducted by infiltration of the *Agrobacteria* suspension into exposed intercellular of the leaves that are still attached to the intact plant. For each recombinant vector construct and control, three plants pot⁻¹ of *Amaranthus* sp. and *Allium porrum* were grown.

In order to have optimum plant growth, *Amaranthus* sp. was grown for up to one month while *Allium porrum* was grown for up to four month. Both crops were irrigated twice in a day at 0800 and 1600. All healthy leaves were chosen for infiltration. The injured leaves were

submerged with the suspension for 1 min. After infiltration the leaves were covered overnight to maintain the humidity prior to transferring back to the glasshouse. Three leaves samples were harvested from each plant at each day three, five and seven. Harvested samples were freeze dried and were stored at -80°C prior to analysis.

2.4. Evaluation of Transient Expression

The GUS staining was performed as previously described (Jefferson, 1987). To compare the transient expression of LHCB with Ubi1P, pieces of leaves from *Amaranthus* sp./*Allium porrum* that was infiltrated with pDRB5b were assayed for GUS analysis to determine the basis for selecting the developmental timing of the mRNA expression of GUS driven by Ubi1P and to measure the extent of the transgene expression in transient systems. The infiltrated leaves were soaked in the GUS staining solution (5 mg of X-Gluc in 0.1 mL of *N,N*-dimethylformamide in 10 mL of buffer containing 50 mM phosphate buffer, 15 mM EDTA, 0.01% Tween-20 pH 7.0). They were incubated for 16 h at 37°C. The stained leaf tissues were rinsed with 70% ethanol until complete decoloration. The pieces of plant leaves infiltrated with pDRB6b (empty vector) served as control plant.

2.5. Data Analysis

All experiments were repeated thrice with at least three replicates for each analysis. The data presented herein are the average results of the experiments and all data were subjected to normality test. One-way Analysis of Variance (ANOVA) was performed for all the raw data using Duncan multiple range tests. All analyses were performed at the level of 5.0% using SPSS 17.0 software package (SPSS Inc., IL, USA).

3. RESULTS

3.1. Recombinant Vector Analysis

The isolated DNA fragments having heterologous ends were successfully inserted in between the RB2-LB of the pDRB6b vector. Our recombinant vector constructs for transient gene expression, p4a11 and p4c9 were designed to contain the partial conserved cDNA sequence of Tocopherol Cyclase gene (TC) from *Elaeis guineensis*. They were controlled by Ubi1P and LHCB, respectively and NosT as the termination sites. All the digested recombinant vector constructs successfully produced the DNA fragments of the expected band size. The PCR product which was

sequenced and subjected to nucleotide blast, shared 98% identity with TC from *Elaeis oleifera* (EU057618.1), 79% *Vitis vinifera* (XM002281388.2), 76% *Glycine max* (XM003522656.1) and *Gossypium hirsutum* (DQ456881.1), 77% to *Ricinus communis* (XM002516502.1) and *Eucalyptus gunnii* (AY336944.1).

3.2. Evaluation of Transient Expression

Reporter gene (GUS) was used to assay for the activity of a particular promoter in a cell or organism. The expression of GUS driven by Ubi1P in pDRB5b showed by the blue staining was visible at day three, five and seven on the leaf of *Amaranthus* sp. while in *Allium porrum* at day five and seven (**Fig. 3a, b**). The results showed that the Ubi1P starts the expression on day three till day seven for *Amaranthus* sp. but on day five till seven for *Allium porrum*. Therefore, this timing data could be used to compare with the expression pattern of LHCB and Ubi1 promoter. However, this result will not elucidate the level of α -tocopherol suppression and α -tocotrienol induction.

3.3. Co-suppression Profile of α -Tocopherol

The HPLC assay was used to determine the concentration of α -tocopherol and α -tocotrienol in the experimental leaves. The retention times were confirmed using standard compounds of α -tocopherol and α -tocotrienol. The results of HPLC analysis are shown in **Table 3**. The *Amaranthus* sp. leaves infiltrated with p4a11 showed significantly reduced concentration ($p \leq 0.05$) of α -tocopherol and that infiltrated with p4c9 also showed lesser concentration of α -tocopherol when compared with the control on all the tested days. In the *Allium porrum* infiltrated with p4a11, significant reduction in α -tocopherol was observed on day seven only. This was tally with the result obtained for GUS analysis. The other tested days showed insignificant reduction when compared with that of the control. In addition, in the *Allium porrum* infiltrated with p4c9 showed significant losses on day three and seven while insignificant losses were detected on day five compared to the control. Interestingly, in all the extracts with suppressed α -tocopherol production a peak corresponding to α -tocotrienol was detected on the HPLC chromatogram (data not shown). It could be seen from **Table 3** that a significant induced production of α -tocotrienol was detected in both *Amaranthus* sp. and *Allium porrum*.

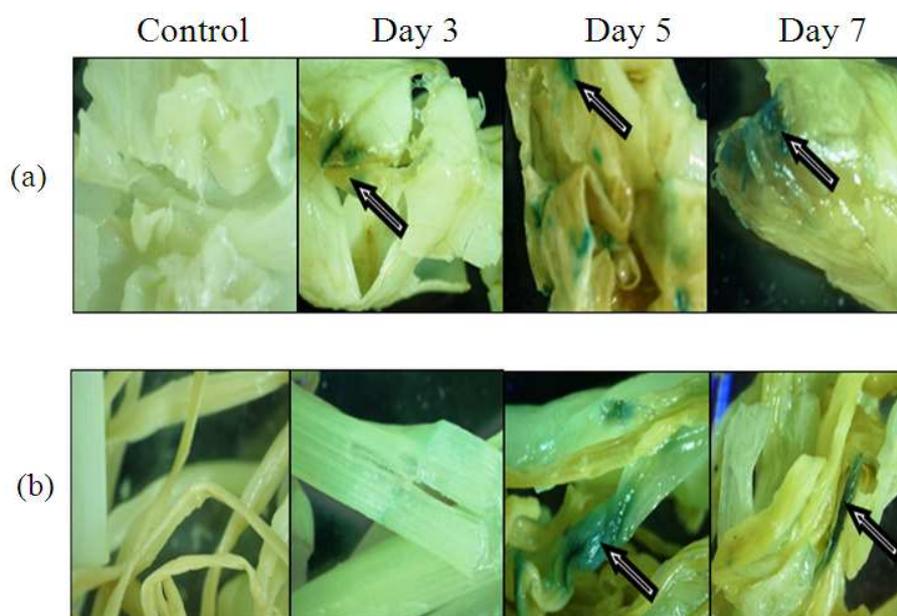


Fig. 3. Histochemical GUS staining. The infiltrated plant leaves with pDRB5b vector were incubated at 37°C for 16 h. The expression of GUS driven by Ubi1P showed by the blue staining (arrow) was highly visible only at day three, five and seven for (a) *Amaranthus* sp. and low visibility on day three and high visibility on day five and seven for (b) *Allium porrum*

Table 3. Summary of the HPLC data on α -tocochromanol concentration and percentage of α -tocopherol suppression after transient transformation by p4a11 and p4c9 compared to control pDRB6b on day three, five and seven

		α -tocopherol (nmol g ⁻¹ tissue)		% α -tocopherol suppression		α -tocotrienol (nmol g ⁻¹ tissue)	
		-----		-----		-----	
Days		<i>Amaranthus</i> sp.	<i>Allium porrum</i>	<i>Amaranthus</i> sp.	<i>Allium porrum</i>	<i>Amaranthus</i> sp.	<i>Allium porrum</i>
Control	3	69+13 ^a	17+0 ⁱ	0	0	0	0
	5	74+11 ^c	25+1 ^k	0	0	0	0
	7	45+8 ^f	26+2 ^l	0	0	0	0
p4a11	3	10+5 ^b	15+2 ⁱ	85	11	3+2 ^o	0 ^u
	5	22+5 ^d	23+1 ^k	71	7	10+1 ^p	16+1.34 ^v
	7	12+3 ^g	18+1 ^m	74	29	13+1.50 ^q	24+2.63 ^w
p4c9	3	0 ^b	9+0 ^j	100	45	7+4 ^r	31+2 ^x
	5	0 ^e	24+1 ^k	100	4	28+4 ^s	41+2 ^y
	7	0 ^h	11+2 ⁿ	100	57	49+4 ^t	50+2 ^z

The data shown are expressed as mean of three replicates + SE from three representative experiments. SE indicates standard error bar. Same letter denotes as being not significant ($p \geq 0.05$) by Duncun test

4. DISCUSSION

The present studies revealed an interesting observation that the α -tocopherol suppression level is higher in *Amaranthus* sp. when compared with that of *Allium porrum*. This is due to the ability of LHCB

(tissue specific promoter) to accumulate large amounts of the aberrant mRNA of TC than that of Ubi1P (constitutive promoter) that led to higher level of α -tocopherol suppression in *Amaranthus* sp. when compared to the *Allium porrum*. Many studies have suggested that the transgene expression controlled by

a tissue-specific promoter worked more effectively in the specific tissue compared to a constitutive promoter (Jones and Sparks, 2009; Schillberg *et al.*, 2005; Masani Abdul *et al.*, 2009). It also appears to be inconsistent with previous reports where Ubi1P was often used to drive the expression of genes in monocotyledons.

The above discussions confirm that the LHCB had driven more effective expression of TC aberrant mRNA than Ubi1P. Similar observation has been reported by Porfirova *et al.* (2002) who have confirmed that the reduction in TC activity due to gene silencing will result in the loss of α -tocopherol. Besides the usage of the promoters, the cause of the different levels of suppression in *Amaranthus* sp. and *Allium porrum* could be due to the different types of photosynthetic pathways in them. *Amaranthus* sp. uses C_4 photosynthetic pathway while *Allium porrum* uses C_3 photosynthetic pathway. The two photosynthetic pathways show significantly different level of gene expression. The *Amaranthus* sp. which is a C_4 plant has reduced state on the shikimate pathway and hence capable of suppressing α -tocopherol easily than C_3 plant (Gowik and Westhoff, 2011). This finding was however different from that reported by Hofius *et al.* (2004) where TC silencing causes severe suppression in α -tocopherol was detected in C_3 plant.

The p4a11 and p4c9 recombinant vector constructs containing partial conserved cDNA of TC gene sequence spanning from nucleotide 45-665 of the orthologous cDNA of EU057618 (Fig. 4) showed over 75% identity with TC of other plant species. This construct enabled the introduction of partial cDNA of the TC to produce the aberrant mRNAs similar to endogenous TC but with distinct lifetime (stabilities). The limited lifetime of aberrant mRNA enables the cell to alter the protein synthesis rapidly in response to its changing needs that lead to the destruction of an mRNA as mRNA lifetimes ranging from several minutes to days. The lesser the stability of a mRNA the lesser the protein will be produced from the particular mRNA. Further investigation of the phenomenon in plants indicated that the down regulation was due to the Post-Transcriptional Gene Silencing (PTGS) of a particular gene due to the presence of a related aberrant gene expression via an increased rate of mRNA degradation (Oka *et al.*, 2010). Therefore, the PTGS mechanism had degraded the population of less stable TC mRNAs

and led to successful suppression of α -tocopherol as the transgene contained higher homology to the cellular TC gene of the endogenous copy which will be degraded too. This shows that the introduced TC gene is sufficient for suppression of endogenous TC transcript level. This phenomenon was also called co-suppression of gene expression or gene silencing (Cameron and Jennings, 1991). According to a study that was previously reported (Abbasi *et al.*, 2007), total blockage of α -tocopherol was achieved through silencing of γ -TMT. But in this study, similar blockage as reported by Porfirova *et al.* (2002) was noticed by silencing TC gene in *Amaranthus* sp. infiltrated with p4c9.

The breakthrough in this study was achieved when the production of α -tocotrienol in infiltrated *Amaranthus* sp./*Allium porrum* was detected parallelly with α -tocopherol suppression and the observation was similar to that reported by Asensi-Fabado and Munne-Bosch (2010). Furthermore, α -tocotrienol production appeared not to be entirely dependent on the level of α -tocopherol suppression as α -tocotrienol produced more in plants infiltrated with p4a11 than p4c9, even though higher suppression of α -tocopherol was detected in p4c9. In addition, α -tocotrienol production was observed in *Allium porrum* higher than in *Amaranthus* sp. even though high suppression was detected in *Amaranthus* sp. In addition, it has been reported that plants which do not accumulate α -tocotrienol may not have a tocotrienol-specific prenyltransferase (HGGT) (Falk and Munne-Bosch, 2010). But, the present study raises an interesting point as to how the silencing of TC could lead to the induction of α -tocotrienol. There are many supporting publications to answer this question. The suppression of α -tocopherol appeared to be sufficient to shift the metabolic pathway towards producing α -tocotrienol as an alternative antioxidant in both plants (Abbasi *et al.*, 2007). Therefore, we are suggesting that the α -tocotrienol production was observed in *Amaranthus* sp. and *Allium porrum* due to the natural ability of the plant to adopt for its survival in α -tocopherol deficient condition after transgenic manipulation of the biosynthetic pathway (Falk and Munne-Bosch, 2010). These cases demonstrate that plants appeared to be flexible in adapting themselves by altering different genes in different ways for survivability.



Fig. 4. Partial conserved cDNA of TC gene sequences (1528 bp) spanning from nucleotide 45-665 of the orthologous cDNA of EU057618 (shown by red arrow). (a): Isolated gene sequence from *Elaeis guineensis*; (b): Gene sequence from *Elaeis oleifera* clone EoEST-835 TC (EU057618)

Secondly, the result also showed that due to the active and stable characteristics of TC enzyme (Collakova and Della Penna 2001) it is not totally silenced and hence the remaining TC mRNAs are capable of producing a fair amount of α -tocopherol. Therefore, α -tocopherol still can be detected in the plants infiltrated with p4a11. Thirdly, the presence of α -tocotrienol in both plants might be due to the involvement of more than a single gene that is able to function as TC enzyme. For example, Homogentisate Solanesyltransferase (HST) is a gene expressed in the leaves required for the biosynthesis of plastoquinones which catalyses the condensation of Solanesyl 2P (SDP) and HGA, accepts farnesyl 2P and geranylgeranyl 2P as prenyl donors for α -tocotrienol production, thus providing further evidence in support of α -tocotrienol production in the absence of a specific HGGT.

In addition, the enhanced supply of HGA may affect the substrate specificity of prenyltransferases leaking

towards α -tocotrienol synthesis. In such plants, α -tocotrienol can accumulate in the leaves (Sadre *et al.*, 2006; Venkatesh *et al.*, 2006). This showed further evidence that HGGT may have no influence but other factors such as composition of plastidic pools of HGA, GGDP and PDP might have influence in the production of α -tocotrienol (Karunanandaa *et al.*, 2005). Besides that, presence of *sucrose export defective1* gene that is capable of encoding enzyme with the same function as TC (Porfirova *et al.*, 2002) could also be the reason of α -tocotrienol production. Interestingly, this gene represents orthologous single copy gene of TC and which may be capable of producing α -tocotrienol (Hofius *et al.*, 2004).

5. CONCLUSION

We have successfully demonstrated the suppression of α -tocopherol level and induction of α -tocotrienol biosynthesis in *Amaranthus* sp. and *Allium porrum* by

silencing gene expression of the Tocopherol/Tocotrienol Cyclase (TC). The level of α -tocochromanol within groups and between groups reported in this study was estimated based on the HPLC assay. The control plant which was infiltrated with pDRB6b vector (empty vector) was totally incapable of producing α -tocotrienol, so the ability of the transiently transformed plant to produce α -tocotrienol is biologically significant. We have developed the HPLC analytical method to correlate the gene expression to the level of metabolite detection. This was developed towards our objective of modifying the nutrient content (α -tocopherol and α -tocotrienol) in the tested plants where the data on metabolite production was mainly required. The excellent results achieved in the present study have motivated us to advance towards the next level of producing stable tocotrienol-biofortified transformed valuable plants that could serve as the source of chemical antioxidants for industrial applications as well as in therapeutics.

6. ACKNOWLEDGEMENT

The authors would like to thank Dr N.M Upadhyaya from CSIRO Plant Industry for providing a series of pDRB vectors and *A. tumefaciens* (AGL1). The senior author would like to express appreciation to Universiti Malaya for providing financial assistance in the form of Skim Latihan Akademik IPTA scholarship.

7. REFERENCES

- Abbasi, A.R., M. Hajirezaei, D. Hofius, U. Sonnewald and L.M. Voll, 2007. Specific roles of α and γ tocopherol in abiotic stress responses of transgenic tobacco. *Plant Physiol.*, 143: 1720-1738. DOI: 10.1104/pp106.094771
- Ajjawi, I. and D. Shintani, 2004. Engineered plants with elevated vitamin E: A nutraceutical success story. *Trends Biotechnol.*, 22: 104-107. DOI: 10.1016/j.tibtech.2004.01.008
- Asensi-Fabado, M.A. and S. Munne-Bosch, 2010. Vitamins in plants: Occurrence, biosynthesis and antioxidant. *Trends Plant Sci.*, 15: 582-592. DOI:0.1016/j.tplants.2010.07.003
- Cameron, F.H. and P.A. Jennings, 1991. Inhibition of gene expression by a short sense fragment. *Nucleic Acids Res.*, 19: 469-475. PMID: 2011521
- Collakova, E. and D. Della Penna, 2001. Isolation and functional analysis of homogentisate phytyltransferase from *Synechocystis* sp. PCC 6803 and *Arabidopsis*. *Plant Physiol.*, 127: 1113-1124. PMID: 11706191
- Falk, F. and S. Munne-Bosch, 2010. Tocochromanol functions in plants: Antioxidation and beyond. *J. Exp. Botany*, 61: 1549-1566. PMID: 20385544
- Gowik, U. and P. Westhoff, 2011. The path from C₃ to C₄ photosynthesis. *Plant Physiol.*, 155: 56-63. DOI: 10.1104/pp.110
- Hofius, D., M.R. Hajirezaei, M. Geiger, H. Tschiersch and M. Melzer *et al.*, 2004. RNAi-mediated tocopherol deficiency impairs photoassimilate export in transgenic potato plants. *Plant Physiol.*, 135: 1256-1268. DOI: 10.1104/pp.104.043927
- Jefferson, R.A., 1987. Assaying chimeric genes in plants: The GUS gene fusion system. *Plant Molecular Biol. Rep.*, 5: 387-405. DOI: 10.1007/BF02667740
- Jones, H.D. and C.A. Sparks, 2009. Promoter sequences for defining transgene expression. *Methods Molecular Biol.*, 478: 171-184. PMID: 19009446
- Karunanandaa, B., Q. Qi, M. Hao, S.R. Basziz and P.K. Jensen *et al.*, 2005. Metabolically engineered oilseed crops with enhanced seed tocopherol. *Metabolic Eng.* PMID: 16125431
- Lee, M.W. and Y. Yang, 2006. Transient expression assay by agroinfiltration of leaves. *Methods Molecular Biol.*, 9: 223-225. PMID: 16739580
- Sanagi, M.M., A.A. Naim, A. Hussain, S.H. Siregar and M.S. Sarjadi *et al.*, 2006. Development and Application of New Modified Poly (styrene-divinylbenzene) Adsorbents and Chromatography Stationary Phases. 1st Edn., Universiti Teknologi Malaysia, Skudai, Johor, ISBN-10: 9673533229, pp: 113.
- Masani Abdul, M.Y., G.K. Ahmad Parveez, A.M. Dayang Izawati, C.P. Lan and A. Siti Nor Akmar, 2009. Construction of PHB and PHBV multiple-gene vectors driven by an oil palm leaf-specific promoter. *Plasmid*, 62: 191-200. PMID: 19699761
- Napoli, C., C. Lemieux and R. Jorgensen, 1990. Introduction of a chimeric chalcone synthase gene into *Petunia* results in reversible co-suppression of homologous genes in trans. *Plant Cell*, 2: 279-289. PMID: 159885
- Nesaretnam, K., N. Guthrie, A.F. Chambers and K.K. Carrol, 1995. Effects of tocotrienols on the growth of a human breast cancer cell line in culture. *Lipids*, 30: 1139-1143. PMID: 8614304
- Oka, S.I., K. Midorikawa and H. Kodama, 2010. Co suppression and RNAi induced by *Arabidopsis* ortholog gene sequences in tobacco. *Plant Biotechnol. Rep.*, 4: 185-192. DOI: 10.1007/s11816-010-0135-2

- Olimpieri, I., R. Caccia, M.E. Picarella, A. Pucci and E. Santangelo *et al.*, 2011. Constitutive co-suppression of the *GA 20-oxidase1* gene in tomato leads to severe defects in vegetative and reproductive development. *Plant Sci.*, 180: 496-503. PMID: 21421397
- Porfirova, S., E. Bergmuller, S. Tropf, R. Lemke and P. Dormann, 2002. Isolation of an *Arabidopsis* mutant lacking vitamin E and identification of a cyclase essential for all tocopherol biosynthesis. *PNAS*, 99: 12495-12500. PMID: 129473
- Ryynanen, M., A.M. Lampi, P. Salo-Vaananen, V. Ollilainen and V. Piironen, 2004. A small scale sample preparation method with HPLC analysis for determination of tocopherols and tocotrienols in cereals. *J. Food Compos. Analytical*, 17: 749-765. DOI: 10.1016/j.jfca.2003.09.014
- Sadre, R., J. Gruber and M. Frentzen, 2006. Characterization of homogentisate prenyltransferases involved in plastoquinone-9 and tocochromanol biosynthesis. *FEBS Lett.*, 580: 5357-5362. PMID: 16989822
- Sambrook, J., E.F. Fritsch and T. Maniatis, 1989. *Molecular Cloning: A Laboratory Manual*. 2nd Edn., Cold Spring Harbor Laboratory Press, ISBN-10: 0879693096, pp: 3.
- Schillberg, S., R.M. Twyman and R. Fischer, 2005. Opportunities for recombinant antigen and antibody expression in transgenic plants-technology assessment. *Vaccine*, 23: 1764-1769. PMID: 15734038
- Traber, M.G. and J. Atkinson, 2007. Vitamin E, antioxidant and nothing more. *Free Radical Biol. Med.*, 43: 4-15. PMID: 17561088
- Venkatesh, T.V., B. Karunanandaa, D.L. Free, J.M. Rottnek and S.R. Basziz *et al.*, 2006 Identification and characterization of an *Arabidopsis* homogentisate phytyltransferase paralog. *Planta*, 223: 1134-1144. PMID: 16408209