In this study, we examined the ameliorative action of N-Acetylcysteine (NAC) against Titanium Dioxide (TiO2) induced testicular degeneration in albino rats. Adult male albino rats were given saline as a control group, TiO2 (1200 mg kg⁻¹ BW), NAC (100 mg kg⁻¹ BW) and co-treatment of NAC and TiO2 as a protective group for 3 months. Testicular tissues were extracted for changes in testicular gene expression and histopathology. Administration of TiO2 significantly up-regulated mRNA expression of IL-6 and TNF-α and normalized by NAC administration. TiO2 administration downregulated Glutathione-S-Transferase (GST) while increased B-cell Lymphoma2 (BcL2) expressions. Co-administration of rats by NAC together with TiO2 normalized changes in GST and BcL2 expression. Expression of steroidogenesis related genes [Androgen Binding Protein (ABR), 17β-Hydroxysteroid Dehydrogenase (17β-HSD), cytochrome P450 17A (CYP17α) and aromatase] showed down regulation in TiO2 administered groups and normalized when NAC given together with TiO2. Moreover, TiO2 induced toxicity in testes that accompanied by degeneration in seminiferous tubules with congestion, oedema and cell disruption that are partially normalized by co-administration of NAC with TiO2. In conclusion, the present findings confirmed the beneficial effect of NAC to prevent apoptosis in spermatogenic and sertoli cells and testicular dysfunction induced by TiO2 in male albino rats.

Keywords: Titanium Dioxide, N-acetylcysteine, Steroidogenesis Related Enzymes, Gene Expression, Testicular Dysfunction and Apoptosis

1. INTRODUCTION

Titanium Dioxide (TiO2) is widely used as an additive, in paints, in sunscreens and in cosmetics as well as in the environmental decontaminant for air, water and soil by the destruction of pesticides (Fisher and Egerton, 2001; Kaida et al., 2004; Choi et al., 2006; Medina et al., 2007). The rate of cellular damage and oxidative stress of nanoparticles are attributed to the particle size and chemical composition (Hoet et al., 2004). The smaller
the particles of TiO2, the more reactivity, effectiveness and toxicity (Oberdorster, 2006). Oxidative stress induced serious toxicity in body tissues as in lung, kidney, testes, brain and spleen (Li et al., 2010; Sang et al., 2012). Higher doses of TiO2 nanoparticles (324-2592 mg kg⁻¹ bw) are accumulated in various organs as spleen, liver, kidney, lung and testes (Chen et al., 2009). Upon exposure to TiO2, pathological lesions occur in the liver, spleen, kidneys, testes and brain (Shi et al., 2013; Jia et al., 2013). Consequently disruption in organ function occur and alteration in inflammatory and defensive mechanisms of body. Testes are rich in polyunsaturated fatty acids which are the source of ROS. Accumulated data showed that ROS-mediated oxidative stress, alters cellular testes damage (Turner and Lysiak, 2008). ROS regulated antioxidant defense systems through regulation of reduced glutathione activity.

As known, N-Acetylcysteine (NAC) is an antioxidant used extensively as an additive in conditional nutrient (Moschou et al., 2008), NAC acts as a cysteine supplier, maintains or even increases the intracellular levels of glutathione (Stanislawski et al., 2000; 2003; Van de Poll et al., 2006; Sadowska et al., 2007; Atkuri et al., 2007). NAC free thiol group can interacts with electrophilic groups of ROS (Sen and Packer, 2000). It has a protective effect on a wide range of cell types (Cay et al., 2006). Zafarullah et al. (2003) reported that cell growth and survival rate increased as a counteract mechanism to ROS-induced injuries resulted in growth arrest and apoptosis. Thus, NAC can restore the imbalance between pro-oxidant and antioxidant systems during oxidative stress. NAC is safe and well tolerated when administered orally while intravenous administrationof NAC induced some helpath problems (Dodd et al., 2008). Rarely, higher doses of NAC causes skin affection as rashes, fever, headache. It may cause drowsiness, low blood pressure and liver problems (Dodd et al., 2008).

Cytokines are proteins that are secreted and expressed by most of cell types (Feghali and Wright, 1997). They are pharmacologically active with either beneficial or pathologic actions on the target cells. They may pro-inflammatory, inflammatory or anti-inflammatory proteins secreted to maintain body defensive mechanism. Imbalanced expression of cytokines are involved in progression of many diseases (Arend and Gabay, 2004). Their expression reflect the immune and health state of the body. In this study, we investigated the effects of NAC on TiO2 induced alterations in gene expression of steroidogenesis, reduced Glutathione (GST), testicular inflammatory and apoptotic activity (IL-6 and TNF-α and Bcl2 respectively) and testes histopathology and possible protection by NAC in male albino rats. The enzymes of steroidogenesis examined are Androgen Binding Protein (ABR), 17β-hydroxysteroid dehydrogenase (17β-HSD), Cytochrome P450 17A (CYP17α) and aromatase.

2. MATERIALS AND METHODS

2.1. Materials and Animals

Ethidium bromide, agarose and TiO2 anatase form, particle size (25-70 nm) were purchased from Sigma Aldrich Co., MO, USA. The male albino rats were purchased from Egyptian Co for Experimental Animals Import, Helwan, Cairo, Egypt. Gum acacia and NAC were obtained from El-Nasr Co., Cairo, Egypt. Cytokines primers were from MACROGEN, Seoul, Korea. Twenty four male albino rats weighting 150-200 g were obtained from the Animal House in Faculty of Veterinary Medicine, Benha University. Rats gained free access to food and water with 12h/12h dark light cycle. All animals were left one week for adaptation. They were housed in separate well-ventilated cages, under standard conditions.

2.2. Experimental Design

Twenty four rats were divided into 4 groups (6 rats per group). Group I (control group), were kept under standard conditions, fed on balanced diet for 3 months. Group II (NAC treated group) received 100 mg kg⁻¹ body weight NAC once daily by gastric gavage for 3 months. Group III (TiO2 treated group) received 1200 mg kg⁻¹ body weight TiO2 by gavage (1/10 LD 50) in 1 mL of 5% gum acacia solution as a solvent once daily for 3 months. The dose of TiO2 was used based on studies of (Wang et al., 2007). Group IV (NAC+TiO2) received 100 mg kg⁻¹ body weight NAC one hour before TiO2 by gavage once daily for 3 months. At the end of the experiments, the rats were sacrificed. Blood and testes were taken for histopathology and gene expression.

2.3. RNA Extraction and cDNA Synthesis

For preparation of total RNA, testes (approximately 100 mg per sample) were collected from rats, flash frozen in liquid nitrogen and subsequently stored at -70°C in 1 mL Qiazol (QIAGEN...
Inc., Valencia, CA). Frozen samples were homogenized using a Polytron 300 D homogenizer (Brinkman Instruments, Westbury, NY). Chloroform (0.3 mL) were added to the homogenate. After shaking for 30 seconds, the mixture was centrifuged at 4°C and 12,500 rpm for 20 min. The aquatic supernatant layer was transferred and an equal volume of isopropanol was added in new sets of tubes, then shaken for 15 sec and centrifuged at 4°C and 12,500 rpm for 15 min. The pellets of RNA were washed with 70% ethanol and dissolved in Diethylpyrocarbonate (DEPC) water. Electrophoresis was used to check RNA integrity. RNA concentration and purity were determined spectrophotometrically at 260 nm. The ratio of OD (260/280) for all samples was ranged from 1.7 to 1.9.

For cDNA synthesis, 2 µg of total RNA and 0.5 ng oligo dT primer in a total volume of 11 µL sterilized DEPC water were incubated in the PeX 0.5 thermal Cycler (Thermo Electronic Corporation, Milford, Ma) at 65°C for 10 min for denaturation. Then, 4 µL of 5X RT-buffer, 2 µL of 10 mM dNTPs and 100 U Moloney Murine Leukemia Virus (M-MuLV) Reverse Transcriptase (SibEnzyme Ltd. Ak, Novosibirsk, Russia) were added and the total volume was completed up to 20 µL by DEPC water. Finally, the mixture was incubated in the thermal cycler at 37°C for 1 h and at 90°C for 10 min to ensure enzyme inactivation.

2.4. Semi-Quantitative PCR Analysis

Specific primers for our examined genes as shown in table 1 were designed using Oligo-4 computer program and synthesized by Macrogen (Macrogen Company, GAsa-dong, Geumcheon-gu. Korea). PCR reaction (25 µL total volume) was consisted of 1 µL cDNA, 1 µL of 10 picomolar (pM) of each primer (forward and reverse) and 12.5 µL PCR master mix (Promega Corporation, Madison, WI) the volume was brought up to 25 using sterilized, deionized water. PCR was carried out using a PeX 0.5 thermal Cycler at 94°C for for 5 min one cycle, followed by variable cycles (according to degree of gene expression) each of which consisted of denaturation at 94°C for one minute, annealing temperature as seen in table 1 and extension at 72 °C for 1 min, with additional final extension at 72°C for 7 min. As a reference, expression of glyceraldehyde-3-phosphate dDehydrogenase (G3PDH) mRNA was used. PCR products were visualized by electrophorrsis in 1.5% agarose gel (Bio Basic INC. Konrad Cres, Markham Ontario), stained with ethidium bromide in TBE (Tris-Borate-EDTA) buffer. PCR products were photographed using gel documentation system. The intensities of the bands were quantified using ImageJ software (http://imagej.en.softonic.com/).

2.5. Histopathological Examination

The testes of rats at the end of experimetal procedures were collected and fixed in Bouin’s solution and dehydrated in ascending grades of alcohols, cleared in xylene and embedded in paraffin. The samples were casted, then sliced into 5 µm thickness and placed onto glass slides. The slides were stained by general and specific stains (Wilson and Gamble, 2008). PAS and Masson’s Trichrome reactions were carried out based on protocols of Kiernan (1999) and Carson (1990) respectively.

2.6. Statistical Analysis

Results are expressed as means ± S.E of 5 different rats per each group. Statistical analysis was done using ANOVA and Fischer’s post hoc test, with p<0.05 being considered as statistically significant.

| Table 1. PCR conditions for rat cytokines and spermatogenesis genes |
|-------------------------|-------------------------|-------------------------|-------------------------|
| IL-6 (450bp)    | 5'-AGTTGGCTCTTCTTGGGACTGATGT-3' | 5'-TGCTCTGAATTGACCTGCGTTTG-3' | 30 cycles, 57°C 1 min |
| TNF-α (256 bp)  | 5'-CCACCACGCTCTTGTCCTGCTAC-3'  | 5'-ACCACCGTGGTTGCTTTTGCTAC-3' | 30 cycles, 58°C 1 min |
| BeL2 (350 bp)   | 5'-TCC ATT ATA AGC TGT CAC AGA GG -3' | 5'-GGA GAG TTC CTC CAC CAC C -3' | 30 cycles, 55°C 1 min |
| GST (575 bp)    | 5'-GCTGAGGAGGATTTGTTGAAAG-3'  | 5'-GCTGACCACTGCAATGATCAG-3' | 33 cycles, 55°C 1 min |
| ABP (195 bp)    | 5'-CCATTCCCTCTTTGAGGATCAG-3'  | 5'-CAGCTCCACCCCCGTTG-3' | 35 cycles, 65°C 45 sec |
| Aromatase (142 bp) | 5'-GCTCGTCGTCGGAGGATGTTG-3' | 5'-GTGAAATCTGGGGTTGTTG-3' | 33 cycles, 57°C 1 min |
| CYP17 α (138bp) | 5'-GCTAATCTGGAGTGCCATGTTG-3' | 5'-GAGCCGTTAGCTGAGGTTG-3' | 33 cycles, 60°C 1 min |
| 17β-HSD (114 bp) | 5'-GCTCGTCGTCGGAGGATGTTG-3' | 5'-CATGAACTCTGGGTTG-3' | 33 cycles, 53°C 1 min |
| GAPDH (309 bp)  | 5'-AGATCCACACCGATGATTTG-3'  | 5'-TCCCTCAAGATTGTCAGCA-3' | 25 cycles, 52°C 1 min |
3. RESULTS

3.1. IL-6 and TNF-α Expression in Testes after TiO2 and NAC Administration in Male Albino Rats

Administration of TiO2 to rats for 3 months up-regulated IL-6 and TNF-α expression in testes and their expression was normalized in rats administered NAC together with TiO2 (Fig. 1a and b). NAC administration to rats alone has no effect on IL-6 and TNF-α expression.

![Image of RT-PCR expression of IL-6 and TNF-α](image)

3.2. BcL2 and GST Expression in Testes after TiO2 and NAC Administration in Male Albino Rats

To test the antiapoptotic and antioxidant effect of NAC, we tested the expression of BcL2 and GST after administration of TiO2 or NAC either alone or together. Figure 1c and d, show that TiO2 increased BcL2 expression and decreased expression of GST and administration of NAC together with TiO2 normalized such increase in BcL2 expression and the decrease in GST expression.

![Image of RT-PCR expression of BcL2 and GST](image)

**Fig. 1.** RT-PCR expression of IL-6, TNF-α, BcL2 and GST expression after administration of either NAC or TiO2 alone or together in Albino rats. NAC and TiO2 were administered for 3 months as described in materials and methods. RNA was extracted and reverse transcribed (3 µg) and RT-PCR analysis was carried out for of IL-6, TNF-α, BcL2 and GST genes. Densitometric analysis was carried for 3 different rats. *p<0.05 Vs control while # p<0.05 Vs NAC group and $ p<0.05 Vs TiO2.
3.3. Expression of Steroidogenesis Regulating Enzymes in Testis of Albino Rats after TiO2 and NAC Administration

To test the effect of the pollutant on genes of testosterone biosynthesis, rats were treated with and/or without TiO2 in presence or absence of NAC. The expression of ABP, 17β-HSD, CYP17α and aromatase were examined using RT-PCR analysis. As seen in Fig. 2a, TiO2 induced significant decrease in ABP expression but in presence of NAC the suppression was normalized and increased. The down regulation in ABP expression was around 55% of normal expression. Also, NAC inhibited TiO2 induced 70% down regulation in aromatase expression that was normalized in presence of NAC (Fig. 2b). Next, we examined the expression of CYP17α that plays a role in the biosynthesis of steroid hormones. TiO2 induced 43% down regulation in CYP17α expression in testis. Presence of NAC with TiO2 significantly normalized the expression of CYP17α (Fig. 2c). Next, we tested the expression of 17β-HSD, the enzyme responsible for synthesis of testosterone from androstenedione.

![Fig. 2](image)

**Fig. 2.** RT-PCR expression of steroidogenesis related gene expression after administration of either NAC or TiO2 alone or together in Albino rats. NAC and TiO2 were administered for 3 months as described in materials and methods. RNA was extracted and reverse transcribed (3 µg) and RT-PCR expression was done for the changes in ABP, aromatase, CYP17α and 17β-HSD genes. Densitometric analysis was carried for 3 different rats. *p<0.05 Vs control while # p<0.05 Vs NAC group and $ p<0.05 Vs TiO
Fig. 3. Photomicrograph of testes of control, TiO2 and NAC administered rats. In (a), testes characterized by well organized seminiferous tubules (st), which consisted of spermatogonium (g), primary and secondary spermatocytes (s), spermatid(td) and spermatozoa (z); H&E X40. In (b), fine collagen fibres with blood vessels represent the interstitial CT between the seminiferous tubules (arrow). Masson trichrome X40. In (c), the rat testes showed positive PAS reaction in the basement membrane (thick arrow), sertoli cells (•) and spermatozoa (→), while faint PAS reaction was detected in the spermatogenic cells (arrow) PAS X 40. In (d), the testes of the TiO2 administered group showed apoptosis of sertoli cells (arrow) H&E X40. In (e), numerous interstitial oedema spreaded between the seminiferous tubules (o) and apoptosis of germinal layers were also detected (arrow), H&E X40. In (f), some seminiferous tubules showed decrease in the thickness of the germinal layer (arrows) with rare sperms (rz) and interstitial oedema (o) H&E X10. In (g), The testes in NAC and TiO2 administered rats showed partial improvement in the testicular cells while the congestion of the testicular blood vessels (C) and mild oedema (o) still persist PAS X40. In (h), the interstitial oedema was diminished in most of the seminiferous tubules while mild oedema still located in some parts of the interstitial CT (o) and the general architecture of the testes was restored, H&E X40.
As seen in Fig. 2d, 17β-HSD expression is down regulated in TiO2 administered rats and NAC normalized its expression. The overall effect of NAC alone as seen in Fig. 2 is stimulatory for spermatogenesis related gene expression.

3.4. Histopathological Findings in Testes of Albino Rats after TiO2 and NAC Administration

The testes of the control group were consisted of seminiferous tubules surrounded by interstitial connective tissue (CT). The seminiferous tubules lined by spermatogonium, primary and secondary spermatocytes, spermatid and spermatozoa. Pyramidal cells were located between the spermatogenic cells (Sertoli cells). Numerous interstitial cells (Leydig cells) were located in the interstitial CT (Fig. 3a). The interstitial CT mainly consisted of fine collagen fibers. Blood vessels were located between the interstitial CT in the testes of the NAC group (Fig. 3b). Positive PAS reaction was noticed in the basement membrane , sertoli cells and spermatozoa (Fig. 3c), while faint PAS reaction was detected in the spermatogenic cells. The testes of the TiO2 administered group showed apoptosis of sertoli cells (Fig. 3d), numerous interstitial oedema spreaded between the seminiferous tubules and apoptosis of germinal layers were also detected (Fig. 3e), some seminiferous tubules showed decrease in the thickness of the germinal layer with rare sperms (Fig. 3f). The testes in NAC and TiO2 administered rats showed partial improvement in the testicular cells while the congestion of the testicular blood vessels and mild oedema still persist (Fig. 3g), the interstitial oedema was diminished in most of the seminiferous tubules while mild oedema still located in some parts of the interstitial CT and the general architecture of the testes were restored (Fig. 3h).

4. DISCUSSION

In this study, we have reported the importance of NAC on testes function and its antiapoptotic and anti-inflammatory effect and its role in protection of testes. As known, NAC and L-carnitine are highly concentrated in the epididymis and spermatozoa, it serves as an intramitochondrial adjunct for the acyl group in the form of acyl CoA that acts as a substrate for fatty acids oxidation to produce energy for sperm respiration and motility (Kanter et al., 2010). Accumulation of TiO2 in testes down regulated genes responsible for cholesterol transport and steroidogenesis including ABP, P450scc, 17β-HSD, CYP-17α and aromatase. Expression of ABP, CYP17α and 17β-HSD were significantly decreased in rats following exposure to TiO2 for 3 months. These genes play important roles in testosterone biosynthesis, for example, progesterone is converted to 17α-hydroxyprogesterone and androstenedione by CYP17α. Then testosterone is synthesized from androstenedione by 17β-HSD. The decrease in gene expression of CYP17α and 17β-HSD indicates a possible contribution of these genes to the decrease in serum testosterone of male rats.

Apoptosis is a programmed cell death, it affects cell function and viability. BcL2 is known as apoptotic biomarker. Administration of TiO2 for 3 months up-regulated BcL2 expression and NAC down regulate it confirming antiapoptotic activity of NAC as reported by ours and findings of (Aggarwal et al., 2010). Germ cell apoptosis play an important role in normal testicular function (Modi et al., 2003). It is required for normal spermatogenesis and is believed to ensure cellular homeostasis and maintain the balance between germ cells and Sertoli cells (Said et al., 2004). Also, apoptosis regulates spermatogenesis in health and disease status (Kerr, 1992). High rates of apoptosis was recorded in testicular biopsies from infertile men with variable degrees of testicular insufficiency (Jurisicova et al., 1999). Sensitivity to factors modulated apoptosis was higher in the spermatogonia and spermatoocytes, but Sertoli and Leydig cells were comparatively the most resistant (Kanter et al., 2010). Such changes was ameliorated and normalized by NAC administration.

In this study, TiO2 stimulated both oxidative stress that indicated by the decrease in reduced glutathione and inflammatory cytokines (IL-6 and TNF-α). It has been shown that NAC ameliorated the decrease in GST and lipid peroxidation induced by diazion (Oksay et al., 2013). Moreover, the changes in cytokines expression induced by TiO2 are ameliorated by NAC administration. NAC acts as antioxidant that stimulate GSH synthesis (Dodd et al., 2008) and is confirmed in this study by up-regulation of testicular GST expression in TiO2 administered rats. So, NAC may restore the disturbance between pro-oxidant and antioxidant meachisms during oxidative stress and that are coincided with our findings. In the epididymis, free antioxidants (NAC) are taken from circulating blood and actively transported through the epithelial cells into the epididymal plasma, using a specific carrier under regulation of androgens (Enomoto et al., 2002). Moreover, antioxidants especially L-carnitine and NAC act to protect the heart against ischemia/perfusion injuries that lead to myocyte death (Shug et al., 1991).
The protective effect of NAC on testicular damage and dysfunction were explained in studies of Cay et al., 2009; Shi et al., 2013; Jia et al., 2013, which, in turn cause oxidative stress, inflammation and apoptosis resulting in organ injury and failure. Dysfunction of the testes in turn affect reproduction and affects the function of the reproductive system (Jia et al., 2013). The cytochrome P-450 aromatase enzyme, the rate-limiting enzyme in estradiol biosynthesis, it catalyzes the estradiol synthesis from testosterone. The present study demonstrated that aromatase mRNA levels decreased in TiO2 administered rats and NAC normalized aromatase expression in a way to control decreased in TiO2 administered rats and NAC administration has significant upregulation in steroidogenesis gene expression. Co-administration of NAC induced partial improvement in the testicular dysfunction, germinal layer apoptosis and diminished the interstitial oedema in most of the seminiferous tubules induced by TiO2.

5. CONCLUSION

The findings of this study confirmed that co-administration of NAC inhibited harmful effects of TiO2 that disrupted testicular function of male albino rats. NAC has antioxidant and antiapoptotic functions. Moreover, it improved steroidogenesis related gene expression which is ameliorated by TiO2 administration. NAC is a good health supplement for protection of male fertility from environmental toxic biohazards.

6. REFERENCES


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